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(54) Title: GTPASE ASSOCIATED PROTEINS

(57) Abstract

The invention provides human GTPase associated proteins (GTPAP) and polynucleotides which identify and encode GTPAP. The invention also provides expression vectors, host cells, antibodies, agonist, and antagonists. The invention also provides methods for diagnosing, treating, or preventing disorders associated with expression of GTPAP.

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GTPASE ASSOCIATED PROTEINS

TECHNICAL FIELD

This invention relates to nucleic acid and amino acid sequences of GTPase associated proteins and to the use of these sequences in the diagnosis, treatment, and prevention of cell proliferative, autoimmune/inflammatory, and immune system disorders.

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BACKGROUND OF THE INVENTION

Guanine nucleotide binding proteins (GTP-binding proteins) participate in a wide range of regulatory functions in all eukaryotic cells, including metabolism, cellular growth, differentiation, signal transduction, cytoskeletal organization, and intracellular vesicle transport and secretion. In higher organisms they are involved in signaling that regulates such processes as the immune response (Aussel, C. et al (1988) J. Immunol. 140:215-220), apoptosis, differentiation, and cell proliferation including oncogenesis (Dhanasekaran, N. et al. (1998) Oncogene 17:1383-1394). Exchange of bound GDP for GTP followed by hydrolysis of GTP to GDP provides the energy that enables GTP-binding proteins to alter their conformation and interact with other cellular components. The superfamily of GTP-binding proteins consists of several families and may be grouped as translational factors, heterotrimeric GTP-binding proteins involved in transmembrane signaling processes (also called G-proteins), and low molecular weight GTP-binding proteins including the proto-oncogene Ras proteins and products of rab, rap, rho, rac, smg21, smg25, YPT, SEC4, and ARF genes, and tubulins (Kaziro, Y. et al. (1991) Ann. Rev. Biochem. 60:349-400). In all cases, the GTPase activity is regulated through interactions with other proteins.

GTP-binding proteins involved in protein biosynthesis include initiation factor 2 (IF-2), elongation factor 2 (EF-Tu), and elongation factor G (EF-G), observed in prokaryotes; and initiation factor 2 (eIF-2), elongation factor $I\alpha$ (EF- $I\alpha$) and elongation factor 2 (EF-2) observed in eukaryotes (Kaziro, supra). IF-2 promotes the GTP-dependent binding of the tRNA to the small subunit of the ribosome, the step that initiates protein translation. Similarly, elongation factors promote the binding of tRNA and GTP and the displacement of GDP after hydrolysis as protein biosynthesis proceeds.

Heterotrimeric GTP-binding proteins are composed of 3 subunits (α , β and γ) which, in their inactive conformation, associate as a trimer at the inner face of the plasma membrane. G_{α} binds GDP or GTP and contains the GTPase activity. The $\beta\gamma$ complex enhances binding of G_{α} to a receptor. G γ is necessary for the folding and activity of G β . (Neer, E.J. et al. (1994) Nature 371:297-300.) Multiple homologs of each subunit have been identified in mammalian tissues, and different combinations of subunits have specific functions and tissue specificities. (Spiegel, A.M. (1997) J.

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Inher. Metab. Dis. 20:113-121.) G protein activity is triggered by seven-transmembrane cell surface receptors (G-protein coupled receptors) which respond to lipid analogs, amino acids and their derivatives, peptides, cytokines, and specialized stimuli such as light, taste, and odor. Activation of the receptor by its stimulus causes the replacement of the G protein-bound GDP with GTP. G_{α} -GTP dissociates from the receptor/ $\beta\gamma$ complex and each of these separated components can interact with and regulate downstream effectors. The signaling stops when G_{α} hydrolyzes its bound GTP to GDP and reassociates with the $\beta\gamma$ complex (Neer, supra).

The alpha subunits of heterotrimeric G proteins can be divided into four distinct classes. The α -s class is sensitive to ADP-ribosylation by pertussis toxin which uncouples the receptor:G-protein interaction. This uncoupling blocks signal transduction to receptors that decrease cAMP levels which normally regulate ion channels and activate phospholipases. The inhibitory α -I class is also susceptible to modification by pertussis toxin which prevents α -I from lowering cAMP levels. Two novel classes of α subunits refractory to pertussis toxin modification are α -q, which activates phospholipase C, and α -12, which has sequence homology with the Drosophila gene concertina and may contribute to the regulation of embryonic development (Simon, M.I. (1991) Science 252:802-808).

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The mammalian $G\beta$ and $G\gamma$ subunits, each about 340 amino acids long, share more than 80% homology. The GB subunit (also called transducin) contains seven repeating units, each about 43 amino acids long. The activity of both subunits may be regulated by other proteins such as calmodulin and phosducin or the neural protein GAP 43 (D. Clapham and E. Neer, 1993, Nature 365:403-406). The β and γ subunits are tightly associated. The β subunit sequences are highly conserved between species, implying that they perform a fundamentally important role in the organization and function of G-protein linked systems (Van der Voorn L. (1992) Febs. Lett. 307 (2):131-134). They contain seven tandem repeats of the WD-repeat sequence motif, a motif found in many proteins with regulatory functions. WD-repeat proteins contain from four to eight copies of a loosely conserved repeat of approximately 40 amino acids which participates in protein-protein interactions. Mutations and variant expression of β transducin proteins are linked with various disorders. Mutations in LIS1, a subunit of the human platelet activating factor acetylhydrolase, cause Miller-Dieker lissencephaly. RACK1 binds activated protein kinase C, and RbAp48 binds retinoblastoma protein. CstF is required for polyadenylation of mammalian pre-mRNA in vitro and associates with subunits of cleavage-stimulating factor. Defects in the regulation of β -catenin contribute to the neoplastic transformation of human cells. The WD40 repeats of the human F-box protein β TrCP mediate binding to β -catenin, thus regulating the targeted degradation of β -catenin by

ubiquitin ligase (Neer, supra; Hart, M. et al (1999) Curr. Biol. 9:207-210). The γ subunit primary structures are more variable than those of the β subunits. They are often post-translationally modified by isoprenylation and carboxyl-methylation of a cysteine residue four amino acids from the C-terminus; this appears to be necessary for the interaction of the $\beta\gamma$ subunit with the membrane and with other GTP-binding proteins. The $\beta\gamma$ subunit has been shown to modulate the activity of isoforms of adenylyl cyclase, phospholipase C, and some ion channels. It is involved in receptor phosphorylation via specific kinases, and has been implicated in the p21ras-dependent activation of the MAP kinase cascade and the recognition of specific receptors by GTP-binding proteins. (Clapham and Neer, supra).

G-proteins interact with a variety of effectors including adenylyl cyclase (Clapham and Neer, supra). The signaling pathway mediated by cAMP is mitogenic in hormone-dependent endocrine tissues such as adrenal cortex, thyroid, ovary, pituitary, and testes. Cancers in these tissues have been related to a mutationally activated form of a $G\alpha_s$ known as the gsp (Gs protein) oncogene (Dhanasekaran, supra). Another effector is phosducin, a retinal phosphoprotein, which forms a specific complex with retinal $G\beta$ and $G\gamma$ ($G\beta\gamma$) and modulates the ability of $G\beta\gamma$ to interact with retinal $G\alpha$ (Clapham and Neer, supra).

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Irregularities in the GTP-binding protein signaling cascade may result in abnormal activation of leukocytes and lymphocytes, leading to the tissue damage and destruction seen in many inflammatory and autoimmune diseases such as rheumatoid arthritis, biliary cirrhosis, hemolytic anemia, lupus erythematosus, and thyroiditis. Abnormal cell proliferation, including cyclic AMP stimulation of brain, thyroid, adrenal, and gonadal tissue proliferation is regulated by G proteins. Mutations in G_{α} subunits have been found in growth-hormone-secreting pituitary somatotroph tumors, hyperfunctioning thyroid adenomas, and ovarian and adrenal neoplasms (Meij, J.T.A. (1996) Mol. Cell. Biochem. 157:31-38; Aussel, supra).

LMW GTP-binding proteins are GTPases which regulate cell growth, cell cycle control, protein secretion, and intracellular vesicle interaction. They consist of single polypeptides which, like the alpha subunit of the heterotrimeric GTP-binding proteins, are able to bind to and hydrolyze GTP, thus cycling between an inactive and an active state. LMW GTP-binding proteins respond to extracellular signals from receptors and activating proteins by transducing mitogenic signals involved in various cell functions. The binding and hydrolysis of GTP regulates the response of LMW GTP-binding proteins and acts as an energy source during this process (Bokoch, G. M. and Der, C. J. (1993) FASEB J. 7:750-759).

At least sixty members of the LMW GTP-binding protein superfamily have been identified _

and are currently grouped into the ras, rho, arf, sar1, ran, and rab subfamilies. Activated ras genes were initially found in human cancers, and subsequent studies confirmed that ras function is critical in determining whether cells continue to grow or become differentiated. Ras1 and Ras2 proteins stimulate adenylate cyclase (Kaziro, supra), affecting a broad array of cellular processes. Stimulation of cell surface receptors activates Ras which, in turn, activates cytoplasmic kinases. These kinases translocate to the nucleus and activate key transcription factors that control gene expression and protein synthesis (Barbacid, M. (1987) Ann. Rev Biochem. 56:779-827, Treisman, R. (1994) Curr. Opin.Genet. Dev. 4:96-98). Other members of the LMW GTP-binding protein superfamily have roles in signal transduction that vary with the function of the activated genes and the locations of the GTPbinding proteins that initiate the activity. Rho GTP-binding proteins control signal transduction pathways that link growth factor receptors to actin polymerization, which is necessary for normal cellular growth and division. The rab, arf, and sarl families of proteins control the translocation of vesicles to and from membranes for protein processing, localization, and secretion. Vesicle- and target- specific identifiers (v-SNAREs and t-SNAREs) bind to each other and dock the vesicle to the acceptor membrane. The budding process is regulated by the closely related ADP ribosylation factors (ARFs) and SAR proteins, while rab proteins allow assembly of SNARE complexes and may play a role in removal of defective complexes (J. Rothman and F. Wieland (1996) Science 272:227-234). Ran GTP-binding proteins are located in the nucleus of cells and have a key role in nuclear protein import, the control of DNA synthesis, and cell-cycle progression (Hall, A. (1990) Science 249:635-640; Barbacid, M. (1987) Ann. Rev Biochem. 56:779-827; Ktistakis, N. (1998) BioEssays 20:495-504; and Sasaki, T. and Takai, Y. (1998) Biochem. Biophys. Res. Commun. 245:641-645).

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The cycling of LMW GTP-binding proteins between the GTP-bound active form and the GDP-bound inactive form is regulated by additional proteins. Guanosine nucleotide exchange factors (GEFs) increase the rate of nucleotide dissociation by several orders of magnitude, thus facilitating release of GDP and loading with GTP. The best characterized is the mammalian homologue of the Drosophila Son-of-Sevenless protein. Certain Ras-family proteins are also regulated by guanine nucleotide dissociation inhibitors (GDIs), which inhibit GDP dissociation. The intrinsic rate of GTP hydrolysis of the LMW GTP-binding proteins is typically very slow, but it can be stimulated by several orders of magnitude by GTPase-activating proteins (GAPs) (Geyer, M. and Wittinghofer, A. (1997) Curr. Opin. Struct. Biol. 7:786-792). Both GEF and GAP activity may be controlled in response to extracellular stimuli and modulated by accessory proteins such as RalBP1 and POB1. Mutant Ras-family proteins, which bind but can not hydrolyze GTP, are permanently activated, and cause cell proliferation or cancer, as do GEFs that inappropriately activate LMW GTP-binding proteins, such as the human oncogene NET1, a Rho-GEF (Drivas, G. T. et al. (1990) Mol. Cell. Biol.-

10:1793-1798; Alberts, A. S. and Treisman, R. (1998) EMBO J. 14:4075-4085).

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A novel group of GTP-binding proteins is the GTP1/OBG family, which are found in species ranging from bacteria to yeast to humans. These proteins contain characteristic GTP- binding motifs and are similar to one another but do not show sequence homology to other GTP-binding proteins. The exact functions of these proteins are as yet uncertain, but they have been shown to be important for regulation of cell differentiation and development (Okamoto, S. and Ochi, K. (1998). Mol. Microbiol 30:107-119; Sazaka, T. et al. (1992) Biochem. Biophys. Res. Commun. 189:363-370).

The discovery of new GTPase associated proteins and the polynucleotides encoding them satisfies a need in the art by providing new compositions which are useful in the diagnosis, prevention, and treatment of cell proliferative, autoimmune/inflammatory, and immune system disorders.

SUMMARY OF THE INVENTION

The invention features substantially purified polypeptides, GTPase associated proteins,

referred to collectively as "GTPAP" and individually as "GTPAP-1," "GTPAP-2," "GTPAP-3,"

"GTPAP-4," "GTPAP-5," "GTPAP-6," "GTPAP-7," "GTPAP-8," "GTPAP-9," "GTPAP-10,"

"GTPAP-11," "GTPAP-12," "GTPAP-13," "GTPAP-14," "GTPAP-15," "GTPAP-16," "GTPAP-17,"

"GTPAP-18," "GTPAP-19," "GTPAP-20," "GTPAP-21," "GTPAP-22," "GTPAP-23," "GTPAP-24,"

"GTPAP-25," "GTPAP-26," "GTPAP-27," "GTPAP-28," and "GTPAP-29." In one aspect, the

invention provides a substantially purified polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-29 and fragments thereof. The invention also includes a polypeptide comprising an amino acid sequence that differs by one or more conservative amino acid substitutions from an amino acid sequence selected from the group consisting of SEQ ID NO:1-29.

The invention further provides a substantially purified variant having at least 90% amino acid identity to at least one of the amino acid sequences selected from the group consisting of SEQ ID NO:1-29 and fragments thereof. The invention also provides an isolated and purified polynucleotide encoding the polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-29 and fragments thereof. The invention also includes an isolated and purified polynucleotide variant having at least 90% polynucleotide sequence identity to the polynucleotide encoding the polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-29 and fragments thereof.

Additionally, the invention provides an isolated and purified polynucleotide which hybridizes under stringent conditions to the polynucleotide encoding the polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-29 and fragments thereof. The

invention also provides an isolated and purified polynucleotide having a sequence which is complementary to the polynucleotide encoding the polypeptide comprising the amino acid sequence selected from the group consisting of SEQ ID NO:1-29 and fragments thereof.

The invention also provides a method for detecting a polynucleotide in a sample containing nucleic acids, the method comprising the steps of: (a) hybridizing the complement of the polynucleotide sequence to at least one of the polynucleotides of the sample, thereby forming a hybridization complex; and (b) detecting the hybridization complex, wherein the presence of the hybridization complex correlates with the presence of a polynucleotide in the sample. In one aspect, the method further comprises amplifying the polynucleotide prior to hybridization.

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The invention also provides an isolated and purified polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:30-58 and fragments thereof. The invention further provides an isolated and purified polynucleotide variant having at least 90% polynucleotide sequence identity to the polynucleotide sequence selected from the group consisting of SEQ ID NO:30-58 and fragments thereof. The invention also provides an isolated and purified polynucleotide having a sequence which is complementary to the polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:30-58 and fragments thereof.

The invention further provides an expression vector containing at least a fragment of the polynucleotide encoding the polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-29. In another aspect, the expression vector is contained within a host cell.

The invention also provides a method for producing a polypeptide, the method comprising the steps of: (a) culturing the host cell containing an expression vector containing a polynucleotide of the invention under conditions suitable for the expression of the polypeptide; and (b) recovering the polypeptide from the host cell culture.

The invention also provides a pharmaceutical composition comprising a substantially purified polypeptide having the amino acid sequence selected from the group consisting of SEQ ID NO:1-29 and fragments thereof, in conjunction with a suitable pharmaceutical carrier.

The invention further includes a purified antibody which binds to a polypeptide selected from the group consisting of SEQ ID NO:1-29 and fragments thereof. The invention also provides a purified agonist and a purified antagonist to the polypeptide.

The invention also provides a method for treating or preventing a disorder associated with decreased expression or activity of GTPAP, the method comprising administering to a subject in need of such treatment an effective amount of a pharmaceutical composition comprising a substantially

purified polypeptide having the amino acid sequence selected from the group consisting of SEQ ID NO:1-29 and fragments thereof, in conjunction with a suitable pharmaceutical carrier.

The invention also provides a method for treating or preventing a disorder associated with increased expression or activity of GTPAP, the method comprising administering to a subject in need of such treatment an effective amount of an antagonist of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-29 and fragments thereof.

BRIEF DESCRIPTION OF THE TABLES

Table 1 shows polypeptide and nucleotide sequence identification numbers (SEQ ID NOs),

clone identification numbers (clone IDs), cDNA libraries, and cDNA fragments used to assemble fulllength sequences encoding GTPAP.

Table 2 shows features of each polypeptide sequence, including potential motifs, homologous sequences, and methods, algorithms, and searchable databases used for analysis of GTPAP.

Table 3 shows selected fragments of each nucleic acid sequence; the tissue-specific expression patterns of each nucleic acid sequence as determined by northern analysis; diseases, disorders, or conditions associated with these tissues; and the vector into which each cDNA was cloned.

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Table 4 describes the tissues used to construct the cDNA libraries from which cDNA clones encoding GTPAP were isolated.

Table 5 shows the tools, programs, and algorithms used to analyze GTPAP, along with applicable descriptions, references, and threshold parameters.

DESCRIPTION OF THE INVENTION

Before the present proteins, nucleotide sequences, and methods are described, it is understood that this invention is not limited to the particular machines, materials and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

It must be noted that as used herein and in the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same

meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

DEFINITIONS

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"GTPAP" refers to the amino acid sequences of substantially purified GTPAP obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of GTPAP. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of GTPAP either by directly interacting with GTPAP or by acting on components of the biological pathway in which GTPAP participates.

An "allelic variant" is an alternative form of the gene encoding GTPAP. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides. Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

"Altered" nucleic acid sequences encoding GTPAP include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as GTPAP or a polypeptide with at least one functional characteristic of GTPAP. Included within this definition are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding GTPAP, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide sequence encoding GTPAP. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent GTPAP. Deliberate amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of GTPAP is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged

amino acids may include lysine and arginine. Amino acids with uncharged polar side chains having similar hydrophilicity values may include: asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

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The terms "amino acid" and "amino acid sequence" refer to an oligopeptide, peptide, polypeptide, or protein sequence, or a fragment of any of these, and to naturally occurring or synthetic molecules. Where "amino acid sequence" is recited to refer to an amino acid sequence of a naturally occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid sequence.

Amplification is generally carried out using polymerase chain reaction (PCR) technologies well known in the art.

The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of GTPAP. Antagonists may include proteins such as antibodies, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of GTPAP either by directly interacting with GTPAP or by acting on components of the biological pathway in which GTPAP participates.

The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')₂, and Fv fragments, which are capable of binding an epitopic determinant. Antibodies that bind GTPAP polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

The term "antisense" refers to any composition containing a nucleic acid sequence which is

complementary to the "sense" strand of a specific nucleic acid sequence. Antisense molecules may be produced by any method including synthesis or transcription. Once introduced into a cell, the complementary nucleotides combine with natural sequences produced by the cell to form duplexes and to block either transcription or translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" refers to the capability of the natural, recombinant, or synthetic GTPAP, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

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The terms "complementary" and "complementarity" refer to the natural binding of polynucleotides by base pairing. For example, the sequence "5' A-G-T 3" bonds to the complementary sequence "3' T-C-A 5'." Complementarity between two single-stranded molecules may be "partial," such that only some of the nucleic acids bind, or it may be "complete," such that total complementarity exists between the single stranded molecules. The degree of complementarity between nucleic acid strands has significant effects on the efficiency and strength of the hybridization between the nucleic acid strands. This is of particular importance in amplification reactions, which depend upon binding between nucleic acid strands, and in the design and use of peptide nucleic acid (PNA) molecules.

A "composition comprising a given polynucleotide sequence" and a "composition comprising a given amino acid sequence" refer broadly to any composition containing the given polynucleotide or amino acid sequence. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotide sequences encoding GTPAP or fragments of GTPAP may be employed as hybridization probes. The probes may be stored in freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

"Consensus sequence" refers to a nucleic acid sequence which has been resequenced to resolve uncalled bases, extended using the XL-PCR kit (Perkin-Elmer, Norwalk CT) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from the overlapping sequences of one or more Incyte Clones and, in some cases, one or more public domain ESTs, using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (GCG, Madison WI). Some sequences have been both extended and assembled to produce the consensus sequence.

"Conservative amino acid substitutions" are those substitutions that, when made, least interfere with the properties of the original protein, i.e., the structure and especially the function of the

protein is conserved and not significantly changed by such substitutions. The table below shows amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative amino acid substitutions.

	Original Residue	Conservative Substitution
5	Ala	Gly, Ser
	Arg	His, Lys
	Asn	Asp, Gln, His
	Asp	Asn, Glu
	Cys	Ala, Ser
10	Gln	Asn, Glu, His
	Glu	Asp, Gln, His
	Gly	. Ala
	His	Asn, Arg, Gln, Glu
	Ile	Leu, Val
15	Leu	Ile, Val
	Lys	Arg, Gln, Glu
	Met	Leu, Ile
	Phe	His, Met, Leu, Trp, Tyr
	Ser	Cys, Thr
20	Thr	Ser, Val
	Trp	Phe, Tyr
	Tyr	His, Phe, Trp
	Val	Ile, Leu, Thr

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Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the absence of one or more amino acid residues or nucleotides.

The term "derivative" refers to the chemical modification of a polypeptide sequence, or a polynucleotide sequence. Chemical modifications of a polynucleotide sequence can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "fragment" is a unique portion of GTPAP or the polynucleotide encoding GTPAP which is identical in sequence to but shorter in length than the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from 5 to 1000 contiguous nucleotides or amino acid residues. A fragment

used as a probe, primer, antigen, therapeutic molecule. or for other purposes, may be at least 5, 10, 15, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a molecule. For example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50% of a polypeptide) as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

A fragment of SEQ ID NO:30-58 comprises a region of unique polynucleotide sequence that specifically identifies SEQ ID NO:30-58, for example, as distinct from any other sequence in the same genome. A fragment of SEQ ID NO:30-58 is useful, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:30-58 from related polynucleotide sequences. The precise length of a fragment of SEQ ID NO:30-58 and the region of SEQ ID NO:30-58 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

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A fragment of SEQ ID NO:1-29 is encoded by a fragment of SEQ ID NO:30-58. A fragment of SEQ ID NO:1-29 comprises a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-29. For example, a fragment of SEQ ID NO:1-29 is useful as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-29. The precise length of a fragment of SEQ ID NO:1-29 and the region of SEQ ID NO:1-29 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

The term "similarity" refers to a degree of complementarity. There may be partial similarity or complete similarity. The word "identity" may substitute for the word "similarity." A partially complementary sequence that at least partially inhibits an identical sequence from hybridizing to a target nucleic acid is referred to as "substantially similar." The inhibition of hybridization of the completely complementary sequence to the target sequence may be examined using a hybridization assay (Southern or northern blot, solution hybridization, and the like) under conditions of reduced stringency. A substantially similar sequence or hybridization probe will compete for and inhibit the binding of a completely similar (identical) sequence to the target sequence under conditions of reduced stringency. This is not to say that conditions of reduced stringency are such that non-specific binding is permitted, as reduced stringency conditions require that the binding of two sequences to one another be a specific (i.e., a selective) interaction. The absence of non-specific binding may be tested by the use of a second target sequence which lacks even a partial degree of complementarity (e.g., less than about 30% similarity or identity). In the absence of non-specific binding, the

substantially similar sequence or probe will not hybridize to the second non-complementary target sequence.

The phrases "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of residue matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program. This program is part of the LASERGENE software package, a suite of molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in Higgins, D.G. and P.M. Sharp (1989) CABIOS 5:151-153 and in Higgins, D.G. et al. (1992) CABIOS 8:189-191. For pairwise alignments of polynucleotide sequences, the default parameters are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted" residue weight table is selected as the default. Percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polynucleotide sequence pairs.

Alternatively, a suite of commonly used and freely available sequence comparison algorithms is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at http://www.ncbi.nlm.nih.gov/BLAST/. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2 Sequences" can be accessed and used interactively at http://www.ncbi.nlm.nih.gov/gorf/bl2.html. The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

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Reward for match: 1

Penalty for mismatch: -2

Open Gap: 5 and Extension Gap: 2 penalties

Gap x drop-off: 50

Expect: 10

Word Size: 11

Filter: on

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Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the hydrophobicity and acidity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table. As with polynucleotide alignments, the percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polypeptide sequence pairs.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) with blastp set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50

Expect: 10

Word Size: 3

Filter: on

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Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size, and which contain all of the elements required for stable mitotic chromosome segregation and maintenance.

The term "humanized antibody" refers to antibody molecules in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of identity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and may be consistent among hybridization experiments, whereas wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about 1% (w/v) SDS, and about 100 µg/ml denatured salmon sperm DNA.

Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Generally, such wash temperatures are selected to be about 5° C to 20° C lower than the thermal melting point (T_{m}) for the specific sequence at a defined ionic strength and pH. The T_{m} is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating T_{m} and conditions for nucleic acid hybridization are well known and can be found in Sambrook et al., 1989, Molecular Cloning: A Laboratory Manual, 2^{nd} ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY;

specifically see volume 2, chapter 9.

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High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68°C in the presence of about 0.2 x SSC and about 0.1% SDS, for 1 hour. Alternatively, temperatures of about 65°C, 60°C, 55°C, or 42°C may be used. SSC concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, denatured salmon sperm DNA at about 100-200 µg/ml. Organic solvent, such as formamide at a concentration of about 35-50% v/v, may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acid sequences by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., $C_0 t$ or $R_0 t$ analysis) or formed between one nucleic acid sequence present in solution and another nucleic acid sequence immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

The words "insertion" and "addition" refer to changes in an amino acid or nucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect cellular and systemic defense systems.

The term "microarray" refers to an arrangement of distinct polynucleotides on a substrate.

The terms "element" and "array element" in a microarray context, refer to hybridizable polynucleotides arranged on the surface of a substrate.

The term "modulate" refers to a change in the activity of GTPAP. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of GTPAP.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with the second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Generally, operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition.

PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

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"Probe" refers to nucleic acid sequences encoding GTPAP, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acid sequences. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule. Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. "Primers" are short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and identification) of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR).

Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences. Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the specification, including the tables, figures, and Sequence Listing, may be used.

Methods for preparing and using probes and primers are described in the references, for example Sambrook et al., 1989, Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; Ausubel et al., 1987, Current Protocols in Molecular Biology, Greene Publ. Assoc. & Wiley-Intersciences, New York NY; Innis et al., 1990, PCR Protocols, A Guide to Methods and Applications, Academic Press, San Diego CA. PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5. 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to

5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

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A "recombinant nucleic acid" is a sequence that is not naturally occurring or has a sequence that is made by an artificial combination of two or more otherwise separated segments of sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook, <u>supra</u>. The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to transform a cell.

Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be use to vaccinate a mammal wherein the recombinant nucleic acid is expressed, inducing a protective immunological response in the mammal.

The term "sample" is used in its broadest sense. A sample suspected of containing nucleic acids encoding GTPAP, or fragments thereof, or GTPAP itself, may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA,-

RNA, or cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

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The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide containing the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or separated, and are at least about 60% free, preferably about 75% free, and most preferably about 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acids or nucleotides by different amino acids or nucleotides, respectively.

"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

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"Transformation" describes a process by which exogenous DNA enters and changes a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type of host cell being transformed and may include, but is not limited to, viral infection, electroporation, heat shock, lipofection, and particle bombardment. The term "transformed" cells includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 95% or at least 98% or greater sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may

have significant identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternate splicing of exons during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the reference molecule. Species variants are polynucleotide sequences that vary from one species to another. The resulting polypeptides generally will have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 95%, or at least 98% or greater sequence identity over a certain defined length of one of the polypeptides.

THE INVENTION

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The invention is based on the discovery of new human GTPase associated proteins (GTPAP), the polynucleotides encoding GTPAP, and the use of these compositions for the diagnosis, treatment, or prevention of cell proliferative, autoimmune/inflammatory, and immune system disorders.

Table 1 lists the Incyte clones used to assemble full length nucleotide sequences encoding GTPAP. Columns 1 and 2 show the sequence identification numbers (SEQ ID NOs) of the polypeptide and nucleotide sequences, respectively. Column 3 shows the clone IDs of the Incyte clones in which nucleic acids encoding each GTPAP were identified, and column 4 shows the cDNA libraries from which these clones were isolated. Column 5 shows Incyte clones and their corresponding cDNA libraries. Clones for which cDNA libraries are not indicated were derived from pooled cDNA libraries. The Incyte clones in column 5 were used to assemble the consensus nucleotide sequence of each GTPAP and are useful as fragments in hybridization technologies.

The columns of Table 2 show various properties of each of the polypeptides of the invention: column 1 references the SEQ ID NO; column 2 shows the number of amino acid residues in each polypeptide; column 3 shows potential phosphorylation sites; column 4 shows potential glycosylation sites; column 5 shows the amino acid residues comprising signature sequences and motifs; column 6 shows homologous sequences as identified by BLAST analysis; and column 7 shows analytical

methods and in some cases, searchable databases to which the analytical methods were applied. The methods of column 7 were used to characterize each polypeptide through sequence homology and protein motifs.

The columns of Table 3 show the tissue-specificity and diseases, disorders, or conditions

associated with nucleotide sequences encoding GTPAP. The first column of Table 3 lists the
nucleotide SEQ ID NOs. Column 2 lists fragments of the nucleotide sequences of column 1. These
fragments are useful, for example, in hybridization or amplification technologies to identify SEQ ID
NO:30-58 and to distinguish between SEQ ID NO:30-58 and related polynucleotide sequences. The
polypeptides encoded by these fragments are useful, for example, as immunogenic peptides. Column

3 lists tissue categories which express GTPAP as a fraction of total tissues expressing GTPAP.
Column 4 lists diseases, disorders, or conditions associated with those tissues expressing GTPAP as a
fraction of total tissues expressing GTPAP. Column 5 lists the vectors used to subclone each cDNA
library. Of particular note is the specific expression of SEQ ID NO:43 in only one library, a human
testis tissue library; the specific expression of SEQ ID NO:49 in only 4 libraries, one of which is
associated with cell proliferation and 3 of which are associated with inflammation; and the specific
expression of SEQ ID NO:40 in only 5 libraries, 3 of which are associated with cell proliferation and
one of which is associated with inflammation.

The columns of Table 4 show descriptions of the tissues used to construct the cDNA libraries from which cDNA clones encoding GTPAP were isolated. Column 1 references the nucleotide SEQ ID NOs, column 2 shows the cDNA libraries from which these clones were isolated, and column 3 shows the tissue origins and other descriptive information relevant to the cDNA libraries in column 2.

The invention also encompasses GTPAP variants. A preferred GTPAP variant is one which has at least about 80%, or alternatively at least about 90%, or even at least about 95% amino acid sequence identity to the GTPAP amino acid sequence, and which contains at least one functional or structural characteristic of GTPAP.

The invention also encompasses polynucleotides which encode GTPAP. In a particular embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:30-58, which encodes GTPAP.

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The invention also encompasses a variant of a polynucleotide sequence encoding GTPAP. In particular, such a variant polynucleotide sequence will have at least about 70%, or alternatively at least about 90%, or even at least about 95% polynucleotide sequence identity to the polynucleotide sequence encoding GTPAP. A particular aspect of the invention encompasses a variant of a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:30-58 which has at least about 70%, or alternatively at least about 90%, or even at least about

95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting of SEQ ID NO:30-58. Any one of the polynucleotide variants described above can encode an amino acid sequence which contains at least one functional or structural characteristic of GTPAP.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding GTPAP, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring GTPAP, and all such variations are to be considered as being specifically disclosed.

Although nucleotide sequences which encode GTPAP and its variants are generally capable of hybridizing to the nucleotide sequence of the naturally occurring GTPAP under appropriately selected conditions of stringency, it may be advantageous to produce nucleotide sequences encoding GTPAP or its derivatives possessing a substantially different codon usage, e.g., inclusion of non-naturally occurring codons. Codons may be selected to increase the rate at which expression of the peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with which particular codons are utilized by the host. Other reasons for substantially altering the nucleotide sequence encoding GTPAP and its derivatives without altering the encoded amino acid sequences include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced from the naturally occurring sequence.

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The invention also encompasses production of DNA sequences which encode GTPAP and GTPAP derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic sequence may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a sequence encoding GTPAP or any fragment thereof.

Also encompassed by the invention are polynucleotide sequences that are capable of hybridizing to the claimed polynucleotide sequences, and, in particular, to those shown in SEQ ID NO:30-58 and fragments thereof under various conditions of stringency. (See, e.g., Wahl, G.M. and S.L. Berger (1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods Enzymol. 152:507-511.) Hybridization conditions, including annealing and wash conditions, are described in "Definitions."

Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment

of DNA polymerase 1, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Perkin-Elmer), thermostable T7 polymerase (Amersham Pharmacia Biotech, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE amplification system (Life Technologies, Gaithersburg MD). Preferably, sequence preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Perkin-Elmer). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system (Perkin-Elmer), the MEGABACE 1000 DNA sequencing system (Molecular Dynamics, Sunnyvale CA), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art. (See, e.g., Ausubel, F.M. (1997) Short Protocols in Molecular Biology, John Wiley & Sons, New York NY, unit 7.7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853.)

The nucleic acid sequences encoding GTPAP may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a cloning vector. (See, e.g., Sarkar, G. (1993) PCR Methods Applic. 2:318-322.) Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences. (See, e.g., Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186.) A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA. (See, e.g., Lagerstrom, M. et al. (1991) PCR Methods Applic. 1:111-119.) In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art. (See, e.g., Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and PROMOTERFINDER libraries (Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 Primer Analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the template at temperatures of about 68°C to 72°C.

When screening for full-length cDNAs, it is preferable to use libraries that have been

size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence into 5' non-transcribed regulatory regions.

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Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotide-specific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Perkin-Elmer), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotide sequences or fragments thereof which encode GTPAP may be cloned in recombinant DNA molecules that direct expression of GTPAP, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent amino acid sequence may be produced and used to express GTPAP.

The nucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter GTPAP-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

In another embodiment, sequences encoding GTPAP may be synthesized, in whole or in part, using chemical methods well known in the art. (See, e.g., Caruthers, M.H. et al. (1980) Nucleic Acids Symp. Ser. 7:215-223; and Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232.)

Alternatively, GTPAP itself or a fragment thereof may be synthesized using chemical methods. For example, peptide synthesis can be performed using various solid-phase techniques. (See, e.g., Roberge, J.Y. et al. (1995) Science 269:202-204.) Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Perkin-Elmer). Additionally, the amino acid sequence of GTPAP, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof. to produce a variant polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography. (See, e.g., Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421.) The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing. (See, e.g., Creighton, T. (1984) Proteins, Structures and Molecular Properties, WH Freeman, New York NY.)

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In order to express a biologically active GTPAP, the nucleotide sequences encoding GTPAP or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in polynucleotide sequences encoding GTPAP. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of sequences encoding GTPAP. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where sequences encoding GTPAP and its initiation codon and upstream regulatory sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an inframe ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used. (See, e.g., Scharf, D. et al. (1994) Results Probl. Cell Differ. 20:125-162.)

Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding GTPAP and appropriate transcriptional and translational control elements. These methods include <u>in vitro</u> recombinant DNA techniques, synthetic techniques, and <u>in vivo</u> genetic recombination. (See, e.g., Sambrook, J. et al. (1989) <u>Molecular Cloning, A Laboratory Manual</u>, Cold Spring Harbor Press, Plainview NY, ch. 4, 8, and 16-17; Ausubel, F.M. et al. (1995) <u>Current Protocols in Molecular Biology</u>, John Wiley & Sons, New York NY, ch. 9, 13, and 16.)

A variety of expression vector/host systems may be utilized to contain and express sequences encoding GTPAP. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or

tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems. The invention is not limited by the host cell employed.

In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotide sequences encoding GTPAP. For example, routine cloning, subcloning, and propagation of polynucleotide sequences encoding GTPAP can be achieved using a multifunctional E. coli vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid (Life Technologies). Ligation of sequences encoding GTPAP into the vector's multiple cloning site disrupts the *lacZ* gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for in vitro transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence. (See, e.g., Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509.) When large quantities of GTPAP are needed, e.g. for the production of antibodies, vectors which direct high level expression of GTPAP may be used. For example, vectors containing the strong, inducible T5 or T7 bacteriophage promoter may be used.

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Yeast expression systems may be used for production of GTPAP. A number of vectors containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast <u>Saccharomyces cerevisiae</u> or <u>Pichia pastoris</u>. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign sequences into the host genome for stable propagation. (See, e.g., Ausubel, 1995, <u>supra</u>; Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; and Scorer, C.A. et al. (1994) Bio/Technology 12:181-184.)

Plant systems may also be used for expression of GTPAP. Transcription of sequences encoding GTPAP may be driven viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO J. 6:307-311). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used. (See. e.g., Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843: and Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105.) These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. (See, e.g., The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196.)

In mammalian cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, sequences encoding GTPAP may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader

sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain infective virus which expresses GTPAP in host cells. (See, e.g., Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659.) In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.)

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For long term production of recombinant proteins in mammalian systems, stable expression of GTPAP in cell lines is preferred. For example, sequences encoding GTPAP can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched media before being switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be propagated using tissue culture techniques appropriate to the cell type.

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Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine phosphoribosyltransferase genes, for use in *tk* and *apr* cells, respectively. (See, e.g., Wigler, M. et al. (1977) Cell 11:223-232; Lowy, l. et al. (1980) Cell 22:817-823.) Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, *dhfr* confers resistance to methotrexate; *neo* confers resistance to the aminoglycosides neomycin and G-418; and *als* and *pat* confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively. (See, e.g., Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14.) Additional selectable genes have been described, e.g., *trpB* and *hisD*, which alter cellular requirements for metabolites. (See, e.g., Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:8047-8051.) Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), β glucuronidase and its substrate β-glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system.

(See, e.g., Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131.)

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Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding GTPAP is inserted within a marker gene sequence, transformed cells containing sequences encoding GTPAP can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding GTPAP under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

In general, host cells that contain the nucleic acid sequence encoding GTPAP and that express GTPAP may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

Immunological methods for detecting and measuring the expression of GTPAP using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on GTPAP is preferred, but a competitive binding assay may be employed. These and other assays are well known in the art. (See, e.g., Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) Current Protocols in Immunology, Greene Pub. Associates and Wiley-Interscience, New York NY; and Pound, J.D. (1998) Immunochemical Protocols, Humana Press, Totowa NJ.)

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding GTPAP include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide. Alternatively, the sequences encoding GTPAP, or any fragments thereof, may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by Amersham Pharmacia Biotech, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for

ease of detection include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as well as substrates. cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with nucleotide sequences encoding GTPAP may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides which encode GTPAP may be designed to contain signal sequences which direct secretion of GTPAP through a prokaryotic or eukaryotic cell membrane.

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

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In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences encoding GTPAP may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric GTPAP protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of GTPAP activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, c-myc, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metalchelate resins, respectively. FLAG, c-myc, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the GTPAP encoding sequence and the heterologous protein sequence, so that GTPAP may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel (1995, supra, ch. 10).

A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

In a further embodiment of the invention, synthesis of radiolabeled GTPAP may be achieved in vitro using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, ³⁵S-methionine.

Fragments of GTPAP may be produced not only by recombinant means, but also by direct peptide synthesis using solid-phase techniques. (See, e.g., Creighton, supra, pp. 55-60.) Protein synthesis may be performed by manual techniques or by automation. Automated synthesis may be achieved, for example, using the ABI 431A peptide synthesizer (Perkin-Elmer). Various fragments of GTPAP may be synthesized separately and then combined to produce the full length molecule.

THERAPEUTICS

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Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of GTPAP and GTPase associated proteins. In addition, the expression of GTPAP is closely associated with proliferating tissues associated with cancer and fetal development, inflamed tissues, and tissues invovled in the immune response. Therefore, GTPAP appears to play a role in cell proliferative, autoimmune/inflammatory, and immune system disorders. In the treatment of disorders associated with increased GTPAP expression or activity, it is desirable to decrease the expression or activity of GTPAP. In the treatment of disorders associated with decreased GTPAP expression or activity, it is desirable to increase the expression or activity of GTPAP.

Therefore, in one embodiment, GTPAP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of GTPAP. Examples of such disorders include, but are not limited to, a cell proliferative disorder, such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus; an autoimmune/inflammatory disorder, such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis,

autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathycandidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and an immune system disorder, such as acquired immunodeficiency syndrome (AIDS), X-linked agammaglobinemia of Bruton, common variable immunodeficiency (CVI), DiGeorge's syndrome (thymic hypoplasia), thymic dysplasia, isolated IgA deficiency, severe combined immunodeficiency disease (SCID), immunodeficiency with thrombocytopenia and eczema (Wiskott-Aldrich syndrome), Chediak-Higashi syndrome, chronic granulomatous diseases, hereditary angioneurotic edema, and immunodeficiency associated with Cushing's disease, leukemias such as multiple myeloma, and lymphomas such as Hodgkin's disease.

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In another embodiment, a vector capable of expressing GTPAP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of GTPAP including, but not limited to, those described above.

In a further embodiment, a pharmaceutical composition comprising a substantially purified GTPAP in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of GTPAP including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of GTPAP may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of GTPAP including, but not limited to, those listed above.

In a further embodiment, an antagonist of GTPAP may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of GTPAP. Examples of such disorders include, but are not limited to, those cell proliferative, autoimmune/inflammatory, and immune system disorders described above. In one aspect, an antibody which specifically binds GTPAP may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for

bringing a pharmaceutical agent to cells or tissues which express GTPAP.

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In an additional embodiment, a vector expressing the complement of the polynucleotide encoding GTPAP may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of GTPAP including, but not limited to, those described above.

In other embodiments, any of the proteins, antagonists, antibodies, agonists, complementary sequences, or vectors of the invention may be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of GTPAP may be produced using methods which are generally known in the art. In particular, purified GTPAP may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind GTPAP. Antibodies to GTPAP may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are generally preferred for therapeutic use.

For the production of antibodies, various hosts including goats, rabbits, rats, mice, humans, and others may be immunized by injection with GTPAP or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and <u>Corynebacterium parvum</u> are especially preferable.

It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to GTPAP have an amino acid sequence consisting of at least about 5 amino acids, and generally will consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are identical to a portion of the amino acid sequence of the natural protein and contain the entire amino acid sequence of a small, naturally occurring molecule. Short stretches of GTPAP amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to GTPAP may be prepared using any technique which provides for

the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique. (See, e.g., Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; and Cole, S.P. et al. (1984) Mol. Cell Biol. 62:109-120.)

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In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used. (See, e.g., Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; and Takeda, S. et al. (1985) Nature 314:452-454.) Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce GTPAP-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries. (See, e.g., Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-10137.)

Antibodies may also be produced by inducing <u>in vivo</u> production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature. (See, e.g., Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299.)

Antibody fragments which contain specific binding sites for GTPAP may also be generated. For example, such fragments include, but are not limited to, F(ab')₂ fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the F(ab')2 fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity. (See, e.g., Huse, W.D. et al. (1989) Science 246:1275-1281.)

Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between GTPAP and its specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering GTPAP epitopes is generally used, but a competitive binding assay may also be employed (Pound, supra).

Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for GTPAP. Affinity is expressed as an

association constant, K_a, which is defined as the molar concentration of GTPAP-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The K_a determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple GTPAP epitopes, represents the average affinity, or avidity, of the antibodies for GTPAP. The K_a determined for a preparation of monoclonal antibodies, which are monospecific for a particular GTPAP epitope, represents a true measure of affinity. High-affinity antibody preparations with K_a ranging from about 10⁹ to 10¹² L/mole are preferred for use in immunoassays in which the GTPAP-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with K_a ranging from about 10⁶ to 10⁷ L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of GTPAP, preferably in active form, from the antibody (Catty, D. (1988) Antibodies, Volume I: A Practical Approach, IRL Press, Washington, DC; Liddell, J.E. and Cryer, A. (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

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The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of GTPAP-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available. (See, e.g., Catty, supra, and Coligan et al. supra.)

In another embodiment of the invention, the polynucleotides encoding GTPAP, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, the complement of the polynucleotide encoding GTPAP may be used in situations in which it would be desirable to block the transcription of the mRNA. In particular, cells may be transformed with sequences complementary to polynucleotides encoding GTPAP. Thus, complementary molecules or fragments may be used to modulate GTPAP activity, or to achieve regulation of gene function. Such technology is now well known in the art, and sense or antisense oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding GTPAP.

Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide sequences to the targeted organ, tissue, or cell population. Methods which are well known to those skilled in the art can be used to construct vectors to express nucleic acid sequences complementary to the polynucleotides

encoding GTPAP. (See, e.g., Sambrook, supra; Ausubel, 1995, supra.)

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Genes encoding GTPAP can be turned off by transforming a cell or tissue with expression vectors which express high levels of a polynucleotide, or fragment thereof, encoding GTPAP. Such constructs may be used to introduce untranslatable sense or antisense sequences into a cell. Even in the absence of integration into the DNA, such vectors may continue to transcribe RNA molecules until they are disabled by endogenous nucleases. Transient expression may last for a month or more with a non-replicating vector, and may last even longer if appropriate replication elements are part of the vector system.

As mentioned above, modifications of gene expression can be obtained by designing complementary sequences or antisense molecules (DNA, RNA, or PNA) to the control, 5', or regulatory regions of the gene encoding GTPAP. Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, may be employed. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature. (See, e.g., Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177.) A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of sequences encoding GTPAP.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes of the invention may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques

for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by <u>in vitro</u> and <u>in vivo</u> transcription of DNA sequences encoding GTPAP. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

Many methods for introducing vectors into cells or tissues are available and equally suitable for use in vivo, in vitro, and ex vivo. For ex vivo therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art. (See, e.g., Goldman, C.K. et al. (1997) Nat. Biotechnol. 15:462-466.)

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Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and monkeys.

An additional embodiment of the invention relates to the administration of a pharmaceutical or sterile composition, in conjunction with a pharmaceutically acceptable carrier, for any of the therapeutic effects discussed above. Such pharmaceutical compositions may consist of GTPAP, antibodies to GTPAP, and mimetics, agonists, antagonists, or inhibitors of GTPAP. The compositions may be administered alone or in combination with at least one other agent, such as a stabilizing compound, which may be administered in any sterile, biocompatible pharmaceutical carrier including, but not limited to, saline, buffered saline, dextrose, and water. The compositions may be administered to a patient alone, or in combination with other agents, drugs, or hormones.

The pharmaceutical compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial,

intramedullary, intrathecal, intraventricular, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

In addition to the active ingredients, these pharmaceutical compositions may contain suitable pharmaceutically-acceptable carriers comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically. Further details on techniques for formulation and administration may be found in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA).

Pharmaceutical compositions for oral administration can be formulated using pharmaceutically acceptable carriers well known in the art in dosages suitable for oral administration. Such carriers enable the pharmaceutical compositions to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions, and the like, for ingestion by the patient.

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Pharmaceutical preparations for oral use can be obtained through combining active compounds with solid excipient and processing the resultant mixture of granules (optionally, after grinding) to obtain tablets or dragee cores. Suitable auxiliaries can be added, if desired. Suitable excipients include carbohydrate or protein fillers, such as sugars, including lactose, sucrose, mannitol, and sorbitol; starch from corn, wheat, rice, potato, or other plants; cellulose, such as methyl cellulose, hydroxypropylmethyl-cellulose, or sodium carboxymethylcellulose; gums, including arabic and tragacanth; and proteins, such as gelatin and collagen. If desired, disintegrating or solubilizing agents may be added, such as the cross-linked polyvinyl pyrrolidone, agar, and alginic acid or a salt thereof, such as sodium alginate.

Dragee cores may be used in conjunction with suitable coatings, such as concentrated sugar solutions, which may also contain gum arabic, talc, polyvinylpyrrolidone, carbopol gel, polyethylene glycol, and/or titanium dioxide, lacquer solutions, and suitable organic solvents or solvent mixtures. Dyestuffs or pigments may be added to the tablets or dragee coatings for product identification or to characterize the quantity of active compound, i.e., dosage.

Pharmaceutical preparations which can be used orally include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a coating, such as glycerol or sorbitol. Push-fit capsules can contain active ingredients mixed with fillers or binders, such as lactose or starches, lubricants, such as talc or magnesium stearate, and, optionally, stabilizers. In soft capsules, the active compounds may be dissolved or suspended in suitable liquids, such as fatty oils, liquid, or liquid polyethylene glycol with or without stabilizers.

Pharmaceutical formulations suitable for parenteral administration may be formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hanks' solution, Ringer's

solution, or physiologically buffered saline. Aqueous injection suspensions may contain substances which increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. Additionally, suspensions of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils, such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate, triglycerides, or liposomes. Non-lipid polycationic amino polymers may also be used for delivery. Optionally, the suspension may also contain suitable stabilizers or agents to increase the solubility of the compounds and allow for the preparation of highly concentrated solutions.

For topical or nasal administration, penetrants appropriate to the particular barrier to be permeated are used in the formulation. Such penetrants are generally known in the art.

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The pharmaceutical compositions of the present invention may be manufactured in a manner that is known in the art, e.g., by means of conventional mixing, dissolving, granulating, dragee-making, levigating, emulsifying, encapsulating, entrapping, or lyophilizing processes.

The pharmaceutical composition may be provided as a salt and can be formed with many acids, including but not limited to, hydrochloric, sulfuric, acetic, lactic, tartaric, malic, and succinic acids. Salts tend to be more soluble in aqueous or other protonic solvents than are the corresponding free base forms. In other cases, the preparation may be a lyophilized powder which may contain any or all of the following: 1 mM to 50 mM histidine, 0.1% to 2% sucrose, and 2% to 7% mannitol, at a pH range of 4.5 to 5.5, that is combined with buffer prior to use.

After pharmaceutical compositions have been prepared, they can be placed in an appropriate container and labeled for treatment of an indicated condition. For administration of GTPAP, such labeling would include amount, frequency, and method of administration.

Pharmaceutical compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example GTPAP or fragments thereof, antibodies of GTPAP, and agonists, antagonists or inhibitors of GTPAP, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be

determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED $_{50}$ (the dose therapeutically effective in 50% of the population) or LD $_{50}$ (the dose lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the therapeutic index, which can be expressed as the LD $_{50}$ /ED $_{50}$ ratio. Pharmaceutical compositions which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED $_{50}$ with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy. Long-acting pharmaceutical compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

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Normal dosage amounts may vary from about 0.1 μ g to 100,000 μ g, up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

DIAGNOSTICS

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In another embodiment, antibodies which specifically bind GTPAP may be used for the diagnosis of disorders characterized by expression of GTPAP, or in assays to monitor patients being treated with GTPAP or agonists, antagonists, or inhibitors of GTPAP. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for GTPAP include methods which utilize the antibody and a label to detect GTPAP in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

A variety of protocols for measuring GTPAP, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of GTPAP expression. Normal or standard values for GTPAP expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, for example, human subjects, with antibody to GTPAP under conditions suitable for complex formation. The amount of standard complex formation may be quantitated by various methods, such as photometric means. Quantities of GTPAP expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, the polynucleotides encoding GTPAP may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotide sequences, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of GTPAP may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of GTPAP, and to monitor regulation of GTPAP levels during therapeutic intervention.

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In one aspect, hybridization with PCR probes which are capable of detecting polynucleotide sequences, including genomic sequences, encoding GTPAP or closely related molecules may be used to identify nucleic acid sequences which encode GTPAP. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the probe identifies only naturally occurring sequences encoding GTPAP, allelic variants, or related sequences.

Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the GTPAP encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:30-58 or from genomic sequences including promoters, enhancers, and introns of the GTPAP gene.

Means for producing specific hybridization probes for DNAs encoding GTPAP include the cloning of polynucleotide sequences encoding GTPAP or GTPAP derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such as ³²P or ³⁵S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotide sequences encoding GTPAP may be used for the diagnosis of disorders associated with expression of GTPAP. Examples of such disorders include, but are not limited to. a cell proliferative disorder, such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis. cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus; an autoimmune/inflammatory disorder, such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis. Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and an immune system disorder, such as acquired immunodeficiency syndrome (AIDS), X-linked agammaglobinemia of Bruton. common variable immunodeficiency (CVI), DiGeorge's syndrome (thymic hypoplasia), thymic dysplasia, isolated IgA deficiency, severe combined immunodeficiency disease (SCID), immunodeficiency with thrombocytopenia and eczema (Wiskott-Aldrich syndrome), Chediak-Higashi syndrome, chronic granulomatous diseases, hereditary angioneurotic edema, and immunodeficiency associated with Cushing's disease, leukemias such as multiple myeloma, and lymphomas such as Hodgkin's disease. The polynucleotide sequences encoding GTPAP may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin. and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered GTPAP expression. Such qualitative or quantitative methods are well known in the art.

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In a particular aspect, the nucleotide sequences encoding GTPAP may be useful in assays that detect the presence of associated disorders, particularly those mentioned above. The nucleotide sequences encoding GTPAP may be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of nucleotide sequences encoding GTPAP in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

In order to provide a basis for the diagnosis of a disorder associated with expression of GTPAP, a normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding GTPAP, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

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Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier thereby preventing the development or further progression of the cancer.

Additional diagnostic uses for oligonucleotides designed from the sequences encoding GTPAP may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced in vitro. Oligomers will preferably contain a fragment of a

polynucleotide encoding GTPAP, or a fragment of a polynucleotide complementary to the polynucleotide encoding GTPAP, and will be employed under optimized conditions for identification of a specific gene or condition. Oligomers may also be employed under less stringent conditions for detection or quantification of closely related DNA or RNA sequences.

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Methods which may also be used to quantify the expression of GTPAP include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves. (See, e.g., Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et al. (1993) Anal. Biochem. 212:229-236.) The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotide sequences described herein may be used as targets in a microarray. The microarray can be used to monitor the expression level of large numbers of genes simultaneously and to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, and to develop and monitor the activities of therapeutic agents.

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Microarrays may be prepared, used, and analyzed using methods known in the art. (See, e.g., Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/251116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. USA 94:2150-2155; and Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662.)

In another embodiment of the invention, nucleic acid sequences encoding GTPAP may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355; Price, C.M. (1993) Blood Rev. 7:127-134; and Trask, B.J. (1991) Trends Genet. 7:149-154.)

Fluorescent in situ hybridization (FISH) may be correlated with other physical chromosome mapping techniques and genetic map data. (See, e.g., Heinz-Ulrich, et al. (1995) in Meyers, supra, pp. 965-968.) Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the

location of the gene encoding GTPAP on a physical chromosomal map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder. The nucleotide sequences of the invention may be used to detect differences in gene sequences among normal, carrier, and affected individuals.

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In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps. Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the number or arm of a particular human chromosome is not known. New sequences can be assigned to chromosomal arms by physical mapping. This provides valuable information to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the disease or syndrome has been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation. (See, e.g., Gatti, R.A. et al. (1988) Nature 336:577-580.) The nucleotide sequence of the subject invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

In another embodiment of the invention, GTPAP, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between GTPAP and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest. (See, e.g., Geysen, et al. (1984) PCT application WO84/03564.) In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with GTPAP, or fragments thereof, and washed. Bound GTPAP is then detected by methods well known in the art. Purified GTPAP can also be coated directly onto plates for use in the aforementioned drug screening techniques. Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding GTPAP specifically compete with a test compound for binding GTPAP. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with GTPAP.

In additional embodiments, the nucleotide sequences which encode GTPAP may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following preferred specific embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications, and publications mentioned above and below, in particular U.S. Ser. Nos. 60/109,592, 60/118,610, and 60/127,990 are hereby expressly incorporated by reference.

EXAMPLES

I. Construction of cDNA Libraries

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RNA was purchased from Clontech or isolated from tissues described in Table 4. Some tissues were homogenized and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a suitable mixture of denaturants, such as TRIZOL (Life Technologies), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA purity. In some cases, RNA was treated with DNase. For most libraries, poly(A+) RNA was isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the UNIZAP vector system (Stratagene) or SUPERSCRIPT plasmid system (Life Technologies), using the

recommended procedures or similar methods known in the art. (See, e.g., Ausubel, 1997, <u>supra</u>, units 5.1-6.6.) Reverse transcription was initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Pharmacia Biotech) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g., PBLUESCRIPT plasmid (Stratagene), PSPORT1 plasmid (Life Technologies), or pINCY (Incyte Pharmaceuticals, Palo Alto CA). Recombinant plasmids were transformed into competent E. coli cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5α, DH10B, or ElectroMAX DH10B from Life Technologies.

II. Isolation of cDNA Clones

Plasmids were recovered from host cells by <u>in vivo</u> excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plus Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

III. Sequencing and Analysis

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cDNA sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Perkin-Elmer) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Pharmacia Biotech or supplied in ABI sequencing kits such as the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Perkin-Elmer). Electrophoretic separation of cDNA sequencing reactions and detection of labeled

polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Molecular Dynamics); the ABI PRISM 373 or 377 sequencing system (Perkin-Elmer) in conjunction with standard ABI protocols and base calling software; or other sequence analysis systems known in the art. Reading frames within the cDNA sequences were identified using standard methods (reviewed in Ausubel, 1997, supra, unit 7.7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example V.

The polynucleotide sequences derived from cDNA sequencing were assembled and analyzed using a combination of software programs which utilize algorithms well known to those skilled in the art. Table 5 summarizes the tools, programs, and algorithms used and provides applicable descriptions, references, and threshold parameters. The first column of Table 5 shows the tools, programs, and algorithms used, the second column provides brief descriptions thereof, the third column presents appropriate references, all of which are incorporated by reference herein in their entirety, and the fourth column presents, where applicable, the scores, probability values, and other parameters used to evaluate the strength of a match between two sequences (the higher the score, the greater the homology between two sequences). Sequences were analyzed using MACDNASIS PRO software (Hitachi Software Engineering, South San Francisco CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence alignments were generated using the default parameters specified by the clustal algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

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The polynucleotide sequences were validated by removing vector, linker, and polyA sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programing, and dinucleotide nearest neighbor analysis. The sequences were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM, and PFAM to acquire annotation using programs based on BLAST, FASTA, and BLIMPS. The sequences were assembled into full length polynucleotide sequences using programs based on Phred, Phrap, and Consed, and were screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length amino acid sequences, and these full length sequences were subsequently analyzed by querying against databases such as the GenBank databases (described above), SwissProt, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, and Hidden Markov Model (HMM)-based protein family databases such as PFAM. HMM is a probabilistic approach which analyzes consensus primary structures of gene

families. (See, e.g., Eddy, S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.)

The programs described above for the assembly and analysis of full length polynucleotide and amino acid sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:30-58. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies were described in The Invention section above.

IV. Northern Analysis

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Table 3.

Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound. (See, e.g., Sambrook, <u>supra</u>, ch. 7; Ausubel, 1995, supra, ch. 4 and 16.)

Analogous computer techniques applying BLAST were used to search for identical or related molecules in nucleotide databases such as GenBank or LIFESEQ (Incyte Pharmaceuticals). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

% sequence identity x % maximum BLAST score

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The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. For example, with a product score of 40, the match will be exact within a 1% to 2% error, and, with a product score of 70, the match will be exact. Similar molecules are usually identified by selecting those which show product scores between 15 and 40, although lower scores may identify related molecules.

The results of northern analyses are reported as a percentage distribution of libraries in which the transcript encoding GTPAP occurred. Analysis involved the categorization of cDNA libraries by organ/tissue and disease. The organ/tissue categories included cardiovascular, dermatologic, developmental, endocrine, gastrointestinal, hematopoietic/immune, musculoskeletal, nervous, reproductive, and urologic. The disease/condition categories included cancer, inflammation, trauma, cell proliferation, neurological, and pooled. For each category, the number of libraries expressing the sequence of interest was counted and divided by the total number of libraries across all categories. Percentage values of tissue-specific and disease- or condition-specific expression are reported in

V. Extension of GTPAP Encoding Polynucleotides

The full length nucleic acid sequences of SEQ ID NO:30-58 were produced by extension of

an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment. One primer was synthesized to initiate 5' extension of the known fragment, and the other primer, to initiate 3' extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68°C to about 72°C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

Selected human cDNA libraries were used to extend the sequence. If more than one extension was necessary or desired, additional or nested sets of primers were designed.

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High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg²⁺, (NH₄)₂SO₄, and β-mercaptoethanol, Taq DNA polymerase (Amersham Pharmacia Biotech), ELONGASE enzyme (Life Technologies), and Pfu DNA polymerase (Stratagene), with the following parameters for primer pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ were as follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C.

The concentration of DNA in each well was determined by dispensing 100 μ l PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5 μ l of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the concentration of DNA. A 5 μ l to 10 μ l aliquot of the reaction mixture was analyzed by electrophoresis on a 1% agarose mini-gel to determine which reactions were successful in extending the sequence.

The extended nucleotides were desalted and concentrated, transferred to 384-well plates, digested with CviJI cholera virus endonuclease (Molecular Biology Research, Madison WI), and sonicated or sheared prior to religation into pUC 18 vector (Amersham Pharmacia Biotech). For shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%) agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector (Amersham

Pharmacia Biotech), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction site overhangs, and transfected into competent <u>E. coli</u> cells. Transformed cells were selected on antibiotic-containing media, individual colonies were picked and cultured overnight at 37°C in 384-well plates in LB/2x carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase (Amersham Pharmacia Biotech) and Pfu DNA polymerase (Stratagene) with the following parameters: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min; Step 5: steps 2, 3, and 4 repeated 29 times; Step 6: 72°C, 5 min; Step 7: storage at 4°C. DNA was quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low DNA recoveries were reamplified using the same conditions as described above. Samples were diluted with 20% dimethysulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers and the DYENAMIC DIRECT kit (Amersham Pharmacia Biotech) or the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Perkin-Elmer).

In like manner, the nucleotide sequences of SEQ ID NO:30-58 are used to obtain 5' regulatory sequences using the procedure above, oligonucleotides designed for such extension, and an appropriate genomic library.

VI. Labeling and Use of Individual Hybridization Probes

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Hybridization probes derived from SEQ ID NO:30-58 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μ Ci of [γ - 32 P] adenosine triphosphate (Amersham Pharmacia Biotech), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Pharmacia Biotech). An aliquot containing 10^7 counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and

compared.

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VII. Microarrays

A chemical coupling procedure and an ink jet device can be used to synthesize array elements on the surface of a substrate. (See, e.g., Baldeschweiler, supra.) An array analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced by hand or using available methods and machines and contain any appropriate number of elements. After hybridization, nonhybridized probes are removed and a scanner used to determine the levels and patterns of fluorescence. The degree of complementarity and the relative abundance of each probe which hybridizes to an element on the microarray may be assessed through analysis of the scanned images.

Full-length cDNAs, Expressed Sequence Tags (ESTs), or fragments thereof may comprise the elements of the microarray. Fragments suitable for hybridization can be selected using software well known in the art such as LASERGENE software (DNASTAR). Full-length cDNAs, ESTs, or fragments thereof corresponding to one of the nucleotide sequences of the present invention, or selected at random from a cDNA library relevant to the present invention, are arranged on an appropriate substrate, e.g., a glass slide. The cDNA is fixed to the slide using, e.g., UV cross-linking followed by thermal and chemical treatments and subsequent drying. (See, e.g., Schena, M. et al. (1995) Science 270:467-470; Shalon, D. et al. (1996) Genome Res. 6:639-645.) Fluorescent probes are prepared and used for hybridization to the elements on the substrate. The substrate is analyzed by procedures described above.

VIII. Complementary Polynucleotides

Sequences complementary to the GTPAP-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring GTPAP. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of GTPAP. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the GTPAP-encoding transcript.

IX. Expression of GTPAP

Expression and purification of GTPAP is achieved using bacterial or virus-based expression

systems. For expression of GTPAP in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the trp-lac (tac) hybrid promoter and the T5 or T7 bacteriophage promoter in conjunction with the lac operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express GTPAP upon induction with isopropyl beta-Dthiogalactopyranoside (IPTG). Expression of GTPAP in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant Autographica californica nuclear polyhedrosis virus (AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding GTPAP by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of cDNA transcription. Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect cells in most cases, or human hepatocytes, in some cases. Infection of the latter requires additional genetic modifications to baculovirus. (See Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945.)

In most expression systems, GTPAP is synthesized as a fusion protein with, e.g., glutathione S-transferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step, affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton enzyme from Schistosoma japonicum, enables the purification of fusion proteins on immobilized glutathione under conditions that maintain protein activity and antigenicity (Amersham Pharmacia Biotech). Following purification, the GST moiety can be proteolytically cleaved from GTPAP at specifically engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-His, a stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN). Methods for protein expression and purification are discussed in Ausubel (1995, supra, ch. 10 and 16). Purified GTPAP obtained by these methods can be used directly in the following activity assay.

X. Demonstration of GTPAP Activity

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The role of GTPAP can be assayed in vitro by monitoring the mobilization of Ca⁺⁺ as part of the signal transduction pathway. (See, e.g., Grynkievicz, G. et al. (1985) J. Biol. Chem. 260:3440; McColl, S. et al. (1993) J. Immunol. 150:4550-4555; and Aussel, C. et al. (1988) J. Immunol. 140-215.) The assay requires preloading neutrophils or T cells with a fluorescent dye such as FURA-2.

Upon binding Ca⁺⁺, FURA-2 exhibits an absorption shift that can be observed by scanning the excitation spectrum between 300 and 400 nm, while monitoring the emission at 510 nm. When the cells are exposed to one or more activating stimuli artificially (i.e., anti-CD3 antibody ligation of the T cell receptor) or physiologically (i.e., by allogeneic stimulation), Ca⁺⁺ flux takes place. Ca⁺⁺ flux results from the release of Ca⁺⁺ from intracellular organelles or from Ca⁺⁺ entry into the cell through activated Ca⁺⁺ channels. This flux can be observed and quantified by assaying the cells in a fluorometer or fluorescence activated cell sorter. Measurements of Ca⁺⁺ flux are compared between cells in their normal state and those preloaded with GTPAP. Increased mobilization attributable to increased GTPAP availability results in increased emission.

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Alternatively, GTPAP activity is measured by quantifying the amount of a non-hydrolyzable GTP analogue, GTPyS, bound over a 10 minute incubation period. Varying amounts of GTPAP are incubated at 30°C in 50mM Tris buffer, pH 7.5, containing 1mM dithiothreitol, 1mM EDTA and 1μ M [35S]GTPyS. Samples are passed through nitrocellulose filters and washed twice with a buffer consisting of 50mM Tris-HCl, pH 7.8, 1mM NaN₃, 10mM MgCl₂, 1mM EDTA, 0.5mM dithiothreitol, 0.01mM PMSF, and 200mM NaCl. The filter-bound counts are measured by liquid scintillation to quantify the amount of bound [35S]GTPyS. GTPAP activity may also be measured as the amount of GTP hydrolysed over a 10 minute incubation period at 37°C. GTPAP is incubated in 50mM Tris-HCl buffer, pH 7.8, containing 1mM dithiothreitol, 2mM EDTA, 10μ M [α -32P]GTP, and 1μ M H-rab protein. GTPase activity is initiated by adding MgCl₂ to a final concentration of 10 mM. Samples are removed at various time points, mixed with an equal volume of ice-cold 0.5mM EDTA, and frozen. Aliquots are spotted onto polyethyleneimine-cellulose thin layer chromatography plates, which are developed in 1M LiCl, dried, and autoradiographed. The signal detected is proportional to GTPAP activity.

Alternatively, GTPAP activity may be demonstrated as the ability to interact with its associated Gα or LMW GTPase in an in vitro binding assay. The candidate GTPases are expressed as fusion proteins with glutathione S-transferase (GST), and purified by affinity chromatography on glutathione-Sepharose. The GTPases are loaded with GDP by incubating 20 mM Tris buffer, pH 8.0, containing 100 mM NaCl, 2 mM EDTA, 5 mM MgCl2, 0.2 mM DTT, 100 μM AMP-PNP and 10 μM GDP at 30°C for 20 minutes. GTPAP is expressed as a FLAG fusion proteins in a baculovirus system. Extracts of these baculovirus cells containing GTPAP-FLAG fusion proteins are precleared with GST beads, then incubated with GST-GTPase fusion proteins. The complexes formed are precipitated by glutathione-Sepharose and separated by SDS-polyacrylamide gel electrophoresis. The separated proteins are blotted onto nitrocellulose membranes and probed with commercially available anti-

FLAG antibodies. GTPAP activity is proportional to the amount of GTPAP-FLAG fusion protein detected in the complex.

XI. Functional Assays

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GTPAP function is assessed by expressing the sequences encoding GTPAP at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression vector containing a strong promoter that drives high levels of cDNA expression. Vectors of choice include pCMV SPORT (Life Technologies) and pCR3.1 (Invitrogen, Carlsbad CA), both of which contain the cytomegalovirus promoter. 5-10 μ g of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2 μ g of an additional plasmid containing sequences encoding a marker protein are co-transfected. Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser optics-based technique, is used to identify transfected cells expressing GFP or CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties. FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; down-regulation of DNA synthesis as measured by decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M.G. (1994) Flow Cytometry, Oxford, New York NY.

The influence of GTPAP on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding GTPAP and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding GTPAP and other genes of interest can be analyzed by northern analysis or microarray techniques.

XII. Production of GTPAP Specific Antibodies

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GTPAP substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., Harrington, M.G. (1990) Methods Enzymol. 182:488-495), or other purification techniques, is used to immunize rabbits and to produce antibodies using standard protocols.

Alternatively, the GTPAP amino acid sequence is analyzed using LASERGENE software (DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art. (See, e.g., Ausubel, 1995, supra, ch. 11.)

Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A peptide synthesizer (Perkin-Elmer) using fmoc-chemistry and coupled to KLH (Sigma-Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity. (See, e.g., Ausubel, 1995, supra.) Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-GTPAP activity by, for example, binding the peptide or GTPAP to a substrate, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

XIII. Purification of Naturally Occurring GTPAP Using Specific Antibodies

Naturally occurring or recombinant GTPAP is substantially purified by immunoaffinity chromatography using antibodies specific for GTPAP. An immunoaffinity column is constructed by covalently coupling anti-GTPAP antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Pharmacia Biotech). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing GTPAP are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of GTPAP (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/GTPAP binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and GTPAP is collected.

XIV. Identification of Molecules Which Interact with GTPAP

GTPAP, or biologically active fragments thereof, are labeled with ¹²⁵I Bolton-Hunter reagent. (See, e.g., Bolton A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled GTPAP, washed, and any wells with labeled GTPAP complex are assayed. Data obtained using different concentrations of GTPAP are used to calculate values for the number, affinity, and association of

GTPAP with the candidate molecules.

Various modifications and variations of the described methods and systems of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with certain embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within the scope of the following claims.

Table

Polypeptide SEO ID NO:	Nucleotide SEQ ID NO:	Clone ID	Library	Fragments
1	30	708398	SYNORAT04	568987X31 (MMLR3DT01), 708398H1, 708398X11, 708398X15, 708398X16, 708398X17, and 708398X21 (SYNORAT04), 2170523F6 (ENDCNOT03), 3374750H1 (CONNTUT05)
2	31	1259937	MENITUT03	(STOMNOT02), 1259937F6 and 1259937H1 (CORPNOT02), 1729248F6 (BRSTTUT08), 23), 3129757F6 (LUNGTUT12), 3268746X15F (SKINNOT04)
Э	32	1452285	PENITUT01	1452285F6 and 1452285H1 (PENITUTO1), 2605011H1 (LUNGTUT07), 3505135H1 (ADRENOT11)
4	33	1812894	PROSTUT12	1812894H1, 1812894X12 and 1809113T6 (PROSTUT12), 1904479F6 (OVARNOT07), 2232535X15F1 and 2232535X18F1(PROSNOT16), 2267486X16C1 (UTRSNOT02), 2508562F6 (CONUTUT01)
ιΩ	34	3074884	BONEUNT01	225362F1 (PANCNOT01), 900707R1 (BRSTTUT03), 1339234F6 (COLNTUT03), 1759046R6 (PITUNOT03), 3074884H1 (BONEUNT01), SBDA02767F1
9	35	3452277	UTRSNON03	1684553F6 (PROSNOT15), 1951534H1 (PITUNOT01), 3452277H1 (UTRSNON03), 4092781T6 (BSCNSZT01), SBFA01413F1, SBFA03044F1, SBFA01805F1
7	36	4203832	BRAITUT29	723394F1 (SYNOOAT01), 862290R1, and 862290T1 (BRAITUT03), 1560918F1 (SPLNNOT04), 3509241H1 (CONCNOT01), 4203832H1 (BRAITUT29)
ω	37	104368	BMARNOT02	104368H1 (BMARNOT02), SAEA03574F1, SAEA01063F1, SAEA00392F1, SAEA02287F1
6	38	1441680	THYRNOT03	1441680F6, 1441680H1, and 1441680T6 (THYRNOT03), 1904222F6 (OVARNOT07), 2477983F6 (SMCANOT01)
10	36	1494955	PROSNON01	965986R1 (BRSTNOT05), 1429037F1 and 1429037T1 (SINTBST01), 1453487F6 (PENITUT01), 1486114H1 (CORPNOT02), 1494955H1 (PROSNON01), 1995426R6 (BRSTTUT03), 2112074X18F1 and 2112348R6 (BRAITUT03)
11	40	1508161	LUNGNOT14	1508161F6 and 1508161H1 (LUNGNOT14), 3334303H1 (BRAIFET01), 4755656H1 (BRAHNOT01)
12	41	1811877	PROSTUT12	493795H1 (HNT2NOT01), 1573136H1 (LNODNOT03), 1811877F6 and 1811877H1 (PROSTUT12), 1825223F6 (LSUBNOT03), 2454143H1 (ENDANOT01), 2651022H1 (BLADTUT08), 3487062H1 (EPIGNOT01), 4536531H1 (OVARNOT12), 4795253H1 (LIVRTUT09), 4854087H1 (TESTNOT10), 4906149H2 (TLYMNOT08), 5196386H1 (LUNLTUT04)

Fragments	1574127F6, 3857867X306F1, and 3857867X313F1 (LNODNOTO3), 1848674H1 (LUNGFETO3), 1877170F6 (LEUKNOTO3), 2695307H1 (UTRSNOT12), 4148654H1 (SINITUTO4), 4984182H1 (HELATXTO5), 5288671H1 (LIVRTUSO2)	2012970H1, 2012970R6, 2012970X11F (TESTNOTO3)	022341F1 (ADENINBO1), 198476R6 (KIDNNOT02), 2254315H1 (OVARTUT01), 2370170F6 (ADRENOT07), 2451278F6 (ENDANOT01)	1	1398541F1 (BRAITUT08), 1662585F	(BKSINCIOS), Z415545H1 (HNT3AZTO1), Z985066H1 (CARGDITO1), 3462702H1 (293TF2TO1)	282552R1, 282552X23, and 282552X7 (CARDNOT01), 889783R1	9н1 (SAAC00359R1.comp, SAAB00136R1, SAAC00330R1	041660R1 (TBLYNOT01), 077378R1 (SYNORAB01), 740028R1 (PANCNOT04),	1924025R6 (BRSTTUT01), 2817769	(BRSTNOT14)	473002F1 and 473002R1 (MMLR1DT01), 690999R6 (LUNGTUT02), 997483R1	1740475R6 (HIPONON01), 2109547H1 (BRAITUT03), 2917557H1	(THYMFET03), 4309528H1 (BRAUNOT01), 4990135H1 (LIVRTUT11)	777588R6 and 777588T6 (COLNNOT05), 3421335H1 (UCMCNOT04)	605761F1, 605761H1, and 605761R6 (BRSTTUT01), 1271131X15	(TESTIUT02), 1516985F1 (PANCTUT01), 1524935H1 (UCMCL5T01),	ı	483862H1 and 483862R1 (HNT2RAT01), 1750781X305F1, 1750781X307D2	264041R6 (HNT2AGT01), 826449R1 (PROSNOT06), 1256777H1 (MENTTUT03)), 4614049H1 (BRAHNOT01)	1557708F6 (BLADTUT04), 1922490R6 (BRSTTUT01), 2198779H1), 2541193F7	LNODNOTO8),	SINWLIUI), SUGASISHI (ARTETULI)
Library	LUNGFET03	TESTNOT03	OVARTUT01	HNT3AZT01			PONSAZT01	-		BRSTNOT14			THYMFET03			UCMCNOT04	BRSTTUT01			HNT2RAT01	MENITUT03		SPLNFET02			
Clone	1848674	2012970	2254315	2415545			2707969			2817769			2917557			3421335	605761			483862	1256777		2198779			
Nucleotide SEQ ID NO:	42	43	₽ ₽	45		-	46			47	•		48			49	50			51	52		53			
Polypeptide SEQ ID NO:	13	14	15	16			17			18			19			20	21			22	23		24			

Polypeptide	Nucleotide	Clone	Library	Fragments
SEQ ID NO:	SEQ ID NO:	ΩI		
25	54	2226116	SEMVNOT01	1662607F6 (BRSTNOT09), 1662607T6 (BRSTNOT09), 2226116F6
				(SEMVNOT01), 2226116H1 (SEMVNOT01), 2930011F6 (TLYMNOT04),
				3015747T6 (MUSCNOT07), 4087670H1 (LIVRNOT06)
26	55	2504472	CONUTUT01	420365F1 (BRSTNOT01), 762246R1 (BRAITUT02), 907754R2 (COLNNOT09),
				1007508H1 (HEALDIT02), 1302342F6 (PLACNOT02), 1913887H1
				(PROSTUT04), 2023822F6 (CONNNOT01), 2023822X11R1 (CONNNOT01),
				2504472H1 (CONUTUT01), 2951618F6 (KIDNFET01)
27	56	3029920	HEARFET02	354846T6 (RATRNOT01), 418533R6 (BRSTNOT01), 935073R1 (CERVNOT01),
				1340722F1 (COLNTUT03), 1416203T6 (BRAINOT12), 1524567F1
				(UCMCL5T01), 1773043H1 (MENTUNON3), 2590310H2 (LUNGNOT22),
				3029920H1 (HEARFET02), 4873053H1 (COLDNOT01), 5687696H1
				(BRAIUNT01)
28	57	3332415	BRAIFET01	118166R1 (MUSCNOT01), 1257348H1 (MENITUT03), 1288237T6
				(BRAINOT111), 1335936F6 (COLNNOT13), 1452268H1 (PENITUT01),
				1996016R6 (BRSTTUT03), 2116665R6 (BRSTTUT02), 2206894F6
				(SINTFET03), 2540063H1 (BONRTUT01), 2808268H1 (BLADTUT08),
·				3086221H1 (HEAONOT03), 3127508H1 (LUNGTUT12), 3295812H1
				(TLYJINT01), 3332415H1 (BRAIFET01), 3604705H1 (LUNGNOT30),
				4821203H1 (PROSTUT17), 4970353H1 (KIDEUNC10), 5055775H1
				(COLATMT01)
29	58	4031536	BRAINOT23	029167X3 (SPLNFET01), 350137R1 (LVENNOT01), 408825X1 (EOSIHET02),
				689446X23 (LUNGTUT02), 1963062R6 (BRSTNOT04), 2288043R6
				(BRAINONO1), 4031536H1 (BRAINOT23)

Table 2

Analytical Methods and Databases	BLAST MOTIFS	BLAST	BLAST MOTIFS PFAM BLOCKS PRINTS	BLAST MOTIFS	BLAST MOTIFS
Homologous Sequences	GTP-binding protein [Mus musculus] g53169	cAMP- regulated Guanine nucleotide exchange factor [Rattus norvegicus]	GTP-binding protein [Rattus norvegicus]	Fos-related antigen [Rattus norvegicus] g1016712 Rabaptin-4 [H. sapiens] g3832516	GTP-binding protein [H. sapiens] g2765411
Signature Sequences, Motifs, and Domains	G524-T531: ATP/GTP- binding site motif		G16-T23: ATP/GTP- binding site motif		G230-T237: ATP/GTP- binding site motif
Potential Glycosylation Sites	N446	N244	N33 N74		
Potential Phosphorylation Sites	T30 S224 T405 S499 T533 S558 S701 T737 T845 S864 S6 T152 T268 T412 T442 T464 T514 T528 T693 S814 S815 S823 T880 Y117 Y842	877 T86 877 S30	S159 S199	T14 S42 T237 S270 S347 S360 T371 T395 T433 S500 T3 S13 S96 T316 S430	T44 T114 T219 T297 S314 S341 S356 T412 T24 S72 T91 T328 T388 T394
. Amino Acid Residues	1002	338	211	516	445
Polypeptide SEQ ID NO:	н	2	m	4	

						·
Analytical Methods and Databases	BLAST	MOTIFS PRINTS BLAST PFAM	MOTIFS BLAST PRINTS	MOTIFS BLAST PRINTS	MOTIFS BLAST	MOTIFS BLAST PFAM
Homologous Sequences	Regulator of G-protein signaling-9 [H. sapiens] g3284012	Putative ras- like protein [H. sapiens] g4092830	Phosducin- like protein [Rattus rattus] g1323727	Similar to WD domain Beta transducin- like protein [C. elegans] g5596646	WS beta- transducin repeat protein [Homo sapiens] g4704417	Putative guanine nucleotide releasing factor [Drosophila affinis]
Signature Sequences, Motifs, and Domains		G31-T38:ATP/GTP- binding site motif	E47-G66, S116-E178, Y188-G272: Phosducin signature	L49-S82: Beta G protein	M294-T308: Beta transducin	K6-E130: Ras Guanine exchange factor
Potential Glycosylation Sites	N73	N130 N181			N76 N92 N231 N289 N378 N421	
Potential Phosphorylation Sites	S174 S202 S289 S29 S305 S323 T434 T11 T147 T197 T198 S270 S273 S371 S397 Y125	S182 S210 S254 S13 T56 S110 S182 S32 T46 S66 S177	S92 T2 T3 Y15 S18 S19 S20 S25 S97 T120 S165 S296 T94 S116 T120 S284	T6 Y57 S82 T91 S112 S187 T231 T257 S309 T6 T81 S132 S157 S210 S241 T462	S420 S94 T107 S118 T167 T179 T308 S390 S39 S58 T78 T113 S129 T160 T167 Y174 T199 S216 S291 T302 T323 T359 T384 S423 T438	S90 T55 T140 S190
Amino Acid Residues	445	281	301	485	447	199
Polypeptide SEQ ID NO:	'	7	ω	o.	10	11

		<u> </u>				
Analytical Methods and Databases	MOTIFS BLAST	MOTIFS BLAST	MOTIFS BLAST	MOTIFS BLAST	MOTIFS BLAST PFAM	MOTIFS BLAST
Homologous Sequences	Transducin- like protein [H. sapiens] g414536	Similar to the beta transducin family [C. elegans] g2315521	Rab7C (small GTP binding protein) [Lotus japonicus] g1370186	ATP(GTP)- binding protein [H. sapiens] g3646 <u>1</u> 30	Similar to probable rabGAP [C. elegans] g3925265	Small GTP- binding protein associated protein [Mus musculus] g725274
Signature Sequences, Motifs, and Domains	L10-I24, M96-L110: Beta transducin	L197-F211: Beta transducin	G23-S30: ATP-GTP binding site	G26-T33: ATP-GTP binding site	F307-S544: Probable rabGAP domain	
Potential Glycosylation Sites		N353 N362 N502		N114 N189 N222		N171 N194 N685
Potential Phosphorylation Sites	S57 S67 S99 T150 T346 S416 S467 S500 T522 T684 S99 T156 S209 S285 T331 T360 T388 T430 T477 T650 T688	T10 S15 T49 S97 S102 S104 S112 S113 S377 S432 S638 T46 S54 S84 S97 T177 S217 T307 S401 S450 S504 T515 S546 T547 S561 Y618	S14	T100 T249 S260 T308 T328 S338 S351 S30 T73 T157 S237 T308	S67 T344 S366 S63 S68 S75 S122 S177 S265 T282 T332 S373 S380 S563 T569 S634 S20 T94 S128 S314 T382 T385 T458 T559	\$262 S \$206 S T586 T 1140 T1 T450 S
Amino Acid Residues	694	654	180	374	649	869
Polypeptide SEQ ID NO:	12	13	14	15	16	17

Analytical Methods and Databases	MOTIFS BLAST	MOTIFS BLAST	MOTIFS BLAST	MOTIFS BLAST	MOTIFS BLAST
Homologous Sequences	Putative GTP- binding protein [C. elegans] g3880615	Putative GTP- binding protein [H. sapiens] g3169010	Kidney injury associated protein HW052 Acc No W86322 ADP-ribosylation factor-like protein 3 [Rattus norvegicus]	Putative WD40 repeat protein [A. thaliana] g4191784	TipD; similar to beta transducin family [D. discoideum]
Signature Sequences, Motifs, and Domains	G29-S36: ATP-GTP binding site	G52-T59: ATP-GTP binding site	G19-T26: ATP-GTP binding site	L323-L337: Beta transducin	L141, L148, L155 L: zipper gene regulatory motif
Potential Glycosylation Sites	N60 N230 N286			N79	N159
Potential Phosphorylation Sites	T325 S115 T133 S232 S275 T336 S22 T221 S232 T320	T197 S3 S5 S9 T14 S132 T197 T285 T553 T40 T56 S160 T189 S261 S582 Y20 Y396 Y419	T60 S73 S90 S99 S73 S193	T10 T24 T93 S122 T243 S263 S270 T305 S317 S325 T357 S372 T379 S100 S170 S223 T227 S285 T348	T184 T76 T137 S139 T161 T174 T183 S213
Amino Acid	396	634	196	446	265
Polypeptide SEQ ID NO:	18	19	20	. 21	22

					
Analytical Methods and Databases	MOTIFS PFAM PRINTS	BLAST MOTIFS PFAM PRINTS	BLAST MOTIFS PFAM BLOCKS PRINTS	BLAST MOTIFS PFAM PRINTS	BLAST MOTIFS PFAM PRINTS
Homologous Sequences		WD-repeat protein [Arabidopsis thaliana] g3924603	Predicted GTP binding protein [C. elegans] g3878629	Predicted WD repeat protein [S. cerevesiae] P42935	GTP-binding protein APD08 [H.sapiens] Accession W75771
Signature Sequences, Motifs, and Domains	G10-T17: ATP/GTP binding site (P- 1oop) A4-S72: Ras domain	N297-D336, P345- D383, G481-Q519: Beta-transducin WD40 repeats	G259-S266:ATP/GTP binding site (P- loop): G113-R433: GTP1/OBG domain	R48-E91, L97-S143, F197 K237, V273- W319, W378-A416, W604 K642, A659- G697: Beta- transducin WD40 repeats	G11-T18, G425-S432: ATP/GTP binding site (P-loop) R6-K187: Ras domain
Potential Glycosylation Sites		NS	N22 N383	N23 N264 N576 N600 N789	N118 N154 N346
Potential Phosphorylation Sites	T55 S111 S127 S148 S171 S14 S94 Y103	S388 T488 S30 S75 T111 S149 S220 S237 T255 S305 S325 T339 T359 S363 S509 S172 T195 S211 T378 T438 T470 Y203	S164 S341 T347 S36 S68 S92 T286 S364	S122 T243 T247 T427 S454 S519 T528 S623 S701 S715 S809 T58 S143 S266 T411 S505 S577 S603 T661 S735 T753 S791 T815	T414 S59 T105 S126 T139 T143 S196 T203 S311 S325 T370 T390 S477 T483 S541 T583 T94 S148 T247 Y160 Y383 Y456
Amino Acid Residues	185	554	434	826	618
Polypeptide SEQ ID NO:	23	24	255		27

Polymentide	Amino	Potential	Potential	Signature	Homologous	Analytical
SEO ID NO:	Acid	Phosphorylation Sites	Glycosylation	Sequences, Motifs,	Sequences	Methods and
	Residues		Sites	and Domains		Databases
28	596	S17 S21 S50 S152 S153		A178-L355: Rho-	Guanine	BLAST
1		T533 S539 T594 S36 S38		family guanine	nucleotide	MOTIFS
		S80 T163 T169 S183 S211		nucleotide exchange	regulatory	PFAM
		T240 S306 T329 T417		factor (RhoGEF)	protein (NET1	BLOCKS
		S457 S508 T545 S45 T64		domain	homologue)	
		S88 T124 S139 S299 S451			(Mus	
		S459 S528 S568 Y180		_	musculus]	
		Y364			g3834631	
29	589	T108 S20 T90 S127 S176	N572	L252-S289, G293-	SEL-10	BLAST
		S278 S467 T521 S522 T189		N329, G333-D369,	[C.elegans]	MOTIFS
		S254 T284 T292 T321 T324		G373-D409, E413-	g2677836	PFAM
		T345 T364 T423 S444 T484		D449, G453-D489,		PRINTS
		T527		G493-D532: Beta-		
				transducin WD40		
•				repeats		
				R160-K206: F-box		
*				domain		

Table 3

1263							
Vector	PSPORT1	pINCY	pincy	pincy	pincy	pincy	pincy
Disease or Condition (Fraction of Total)	Cell Proliferation (0.692) Inflammation (0.372)	Cell Proliferation (0.731) Inflammation (0.219) Neurological (0.049)	Cell Proliferation (0.875) Trauma (0.125)	Cell Proliferation (0.647) Inflammation (0.264)	Cell Proliferation (0.507) Inflammation (0.352)	Cell Proliferation (0.667) Inflammation (0.111) Neurological (0.111)	Cell Proliferation (0.641) Inflammation (0.302) Neurological (0.038)
Tissue Expression (Fraction of Total)	Reproductive (0.256) Nervous (0.154) Gastrointestinal (0.154)	Reproductive (0.268) Cardiovascular (0.146) Nervous (0.146)	Cardiovascular (0.375) Reproductive (0.375) Dermatologic (0.125) Endocrine (0.125)	Reproductive (0.412) Gastrointestinal (0.147) Hematopoietic/Immune (0.147)	Nervous (0.211) Reproductive (0.197) Gastrointestinal (0.169)	Reproductive (0.444) Nervous (0.333) Gastrointestinal (0.111) Urologic (0.111)	Nervous (0.340) Reproductive (0.208) Gastrointestinal (0.151)
Selected Fragments	628-711	1094-1129	652-703	1224-1292	16-65	947-1043	840-1001
Nucleotide Seg ID NO:	30	31	32	33	34	35	36

Nucleotide Seq ID NO:	Selected Fragments	Tissue Expression (Fraction of Total)	Disease or Condition (Fraction of Total)	Vector
37	507-551	Hematopoietic/Immune (0.269) Nervous (0.269) Reproductive (0.154)	Inflammation (0.423) Cell Proliferation (0.269)	PBLUESCRIPT
38	218-262	Cardiovascular (0.357) Nervous (0.214) Gastrointestinal (0.143)	Cell Proliferation (0.572) Inflammation (0.214)	pINCY
39	164-208	Nervous (0.280) Reproductive (0.260) Developmental (0.120)	Cell Proliferation (0.740) Inflammation (0.180)	PSPORT1
40	369-411	Cardiovascular (0.250) Developmental (0.250) Gastrointestinal (0.250)	Cell Proliferation (0.500) Inflammation (0.250)	pINCY
41	272-316	Reproductive (0.392) Gastrointestinal (0.118) Hematopoietic/Immune (0.118)	Cell Proliferation (0.626) Inflammation (0.137)	pINCY
42	664-708	Nervous (0.211) Reproductive (0.211) Cardiovascular (0.158)	Cell Proliferation (0.614) Inflammation (0.281)	pINCY
43	226-270	Reproductive (1.000)	Inflammation (1.000)	PBLUESCRIPT
44	11-55	Reproductive (0.254) Gastrointestinal (0.206) Cardiovascular (0.159)	Cell Proliferation (0.698) Inflammation (0.206)	PSPORT1

Vector	pincy	DINCY	pincy	pincy	PINCY	PSPORT1	PBLUESCRIPT	PINCY
Disease or Condition (Fraction of Total)	Cell Proliferation (0.781) Inflammation (0.234)	Cell Proliferation (0.582) Inflammation (0.235)	Cell Proliferation (0.655) Inflammation (0.211)	Cell Proliferation (0.543) Inflammation (0.272)	Inflammation (1.000)	Cell Proliferation (0.778) Inflammation (0.156)	Cell Proliferation (1.000) Inflammation (0.200)	Cell proliferation (0.565) Inflammation (0.369)
Tissue Expression (Fraction of Total)	Reproductive (0.281) Nervous (0.188) Gastrointestinal (0.156)	Nervous (0.330) Reproductive (0.183) Hematopoietic/Immune (0.122)	Nervous (0.218) Reproductive (0.188) Gastrointestinal (0.158)	Reproductive (0.222) Hematopoietic/Immune (0.160) Nervous (0.160)	<pre>Gastrointestinal (0.333) Hematopoietic/Immune (0.333) Reproductive (0.333)</pre>	Reproductive (0.289) Gastrointestinal (0.133) Hematopoietic/Immune (0.133)	Nervous (0.500) Gastrointestinal (0.200) Cardiovascular (0.100)	Nervous (0.326) Reproductive (0.326) Cardiovascular (0.152)
Selected Fragments	637-681	1016-1060	737-781	469-513	. 226-270	456-500	252-296	60-104
Nucleotide Seq ID NO:	45	. 46	47	48	49		51	52

Nucleotide Seq ID NO:	Selected Fragments	Tissue Expression (Fraction of Total)	Disease or Condition (Fraction of Total)	Vector
53	488-532	Reproductive (0.232) Nervous (0.195) Hematopoietic/Immune (0.146)	Cell proliferation (0.622) Inflammation (0.427)	pINCY
54	.686-730	Reproductive (0.250) Gastrointestinal (0.150) Hematopoietic/Immune (0.150)	Cell proliferation (0.700) Inflammation (0.400)	pINCY
5.5	543-587 1299-1343	Reproductive (0.282) Nervous (0.155) Gastrointestinal (0.146)	Cell proliferation (0.592) Inflammation (0.359)	pINCY
26	345-389 792-836	Nervous (0.268) Reproductive (0.169) Cardiovascular (0.113) Hematopoietic/Immune (0.113)	Cell proliferation (0.606) Inflammation (0.296)	pINCY
57	163-207	Reproductive (0.270) Gastrointestinal (0.189) Nervous (0.156)	Cell proliferation (0.705) Inflammation (0.254)	pINCY
58	381-425 726-770	Nervous (0.317) Reproductive (0.250) Gastrointestinal (0.117)	Cell proliferation (0.450) Inflammation (0.283)	pINCY

Table 4

Nucleotide	Library	Library Description
ID NO:		
30	SYNORAT04	This library was constructed using RNA isolated from the wrist synovial membrane tissue of a 62-year-old female with rheumatoid arthritis.
31	MENITUT03	This library was constructed using RNA isolated from brain meningioma tissue removed from a
		35-year-old female during excision of a cerebral meningeal lesion. Pathology indicated a
		n in the right cerebellopontine angle of the brain. Patient
		hypothyroidism. Family history included myocardial infarction and breast cancer.
32	PENITUT01	This library was constructed using RNA isolated from tumor tissue removed from the penis of a 64-year-old male during penile amputation. Pathology indicated a fungating invasive grade
		4 squamous cell carcinoma involving the inner wall of the foreskin and extending onto the
		u
		coronary artery disease, angina pectoris, gout, and obesity. Family history included
33	PROSTITI 2	This library was constructed using RNA isolated from prostate tumor tissue removed from a
)		during a radical prostatectomy. Pathology indicated an adenocarcinoma
		son grade 2+
		with elevated prostate specific antigen (PSA).
34	BONEUNT01	
		cell line (ATCC HTB-85) derived from an 11-year-old Caucasian female.
35	UTRSNON03	This library was constructed from 6.4 million independent clones from a uterine library.
		RNA for these libraries was isolated from uterine myometrial tissue removed from a 41-year-
		old female during a vaginal hysterectomy with dilation and curettage. The endometrium was
		secretory and contained fragments of endometrial polyps. Benign endo- and ectocervical
		mucosa were identified in the endocervix. Pathology for the associated tumor tissue
		indicated uterine leiomyoma. The normalization and hybridization conditions were adapted
	•	from Soares et al. (Proc.Natl.Acad.Sci. USA (1994) 91:9928).
36	BRAITUT29	ŏ
		parietal lobe of a 43-year-old female during excision of a cerebral meningeal lesion.
		Pathology indicated high grade glioma. Family history included acute myocardial infarction,
		atheresternic coronary arcery arease, beingh hypertension, and hyperingran
37	BMARNOT02	This library was constructed using RNA isolated from the bone marrow of 24 male and female Caucasian donors, 16 to 70 years old. (RNA came from Clontech.)

Table 4 (cont.)

Nucl	Nucleotide SEO ID NO:	Library	Library Description
	38	THYRNOT03	This library was constructed using RNA isolated from thyroid tissue removed from the left thyroid of a 28-year-old Caucasian female during a complete thyroidectomy. Pathology indicated a small nodule of adenomatous hyperplasia present in the left thyroid. Pathology for the associated tumor tissue indicated dominant follicular adenoma, forming a well-encapsulated mass in the left thyroid.
	39	PROSNON01	This normalized library was constructed from 4.4 million independent clones from a prostate library. Starting RNA was made from prostate tissue removed from a 28-yearold Caucasian male who died from a self-inflicted gunshot wound. The normalization and hybridization conditions were adapted from Soares, M.B. et al. (1994) Proc. Natl. Acad. Sci. USA 91:9228-9232, using a longer (19 hour) reannealing hybridization period.
71	40	LUNGNOT14	of a ced canu
	41	PROSTUT12	cated a presen
	42	LUNGFET03	removed from a
·	43	TESTNOT03	This library was constructed using RNA isolated from testicular tissue removed from a 37-year-old Caucasian male, who died from liver disease. Patient history included cirrhosis, jaundice, and liver failure.
	44	OVARTUT01	This library was constructed using RNA isolated from ovarian tumor tissue removed from a 43-year-old Caucasian female during removal of the fallopian tubes and ovaries. Pathology indicated grade 2 mucinous cystadenocarcinoma involving the entire left ovary. Patient history included mitral valve disorder, pneumonia, and viral hepatitis. Family history included atherosclerotic coronary artery disease, pancreatic cancer, stress reaction, cerebrovascular disease, breast cancer, and uterine cancer.
	45	HNT3AZT01	This library was constructed using RNA isolated from the hNT2 cell line (derived from a human teratocarcinoma that exhibited properties characteristic of a committed neuronal precursor). Cells were treated for three days with 0.35 micromolar 5-aza-2'deoxycytidine (AZ).

Table 4 (cont.)

Nucleotide	Library	Library Description
SEQ ID NO:	•	
46	PONSAZT01	This library was constructed using RNA isolated from diseased pons tissue from the brain of a 74-year-old Caucasian male who died from Alzheimer's disease.
47	BRSTNOT14	This library was constructed using RNA isolated from breast tissue obtained from a 62- yearold Caucasian female during a unilateral extended simple mastectomy. Pathology for the
		ř
		hyperlipidemia, cardiac dysrhythmia, and obesity. Family history included atherosclerotic
		coronary artery disease, myocardial intarction, colon cancer, ovarian cancer, lung cancer, and cerebrovascular disease.
48	THYMFET03	This library was constructed using RNA isolated from thymus tissue removed from a Caucasian
		male fetus.
49	UCMCNOT04	This library was constructed using RNA isolated from mononuclear cells obtained from the
		umbilical cord blood of multiple individuals of mixed age and sex. The cells were treated
		With G-CSF.
05 72	BRSTTUT01	This library was constructed using RNA isolated from breast tumor tissue removed from a 55
-	-	indicated invasive grade 4 mammary adenocarcinoma of mixed lobular and ductal type,
		extensively involving the left breast. Family history included benign hypertension,
		atheroscierotic coronary artery disease, cerebrovascular disease, and depressive disorder.
51	HNT2RAT01	Stratagene (STR937231), using RNA isolated from the hNT2
		ell line (derived from a human te
		neuronal precursor). Cells were treated with retinoic acid for 2
. 52	MENITUT03	' was constructed using RNA isolated from brain meningioma tissue r
		ц
	-	
		history included hypothyroidism. Family history included myocardial infarction and breast
		cancer.
. 53	SPLNFET02	This library was constructed using RNA isolated from spleen tissue removed from a Caucasian
54	SEMVNOT01	This library was constructed using RNA isolated from seminal vesicle tissue removed from a
		The state of the s
		oo-year-oid cadcasian male during radical prostatectomy. Facinology for the associated tumor theshe indicated adenocatchoms (Gleason grade 3+2) of the prostate. Adenofibromatons
		hyperplasia was also present. The patient presented with elevated prostate specific antigen
		(PSA). Family history included a malignant breast neoplasm.

Table 4 (cont.)

•		
Nucleotide	Library	Library Description
SEQ ID NO:		
55	CONUTUT01	
		obtained Irom a ol-year-old lemale duling a cocal abdominat hysterecount and areastatic salpingo-ophorectomy with regional lymph node excision. Pathology indicated a metastatic
		grade 4 malignant mixed mullerian tumor present in the sigmoid mesentery at two sites.
56	HEARFET02	HEARFET02 This library was constructed using RNA isolated from heart tissue removed from a Caucasian
		male fetus, who was stillborn at 23 weeks' gestation with a hypoplastic left heart.
57	BRAIFET01	This library was constructed using RNA isolated from brain tissue removed from a Caucasian
		male fetus, who was stillborn at 23 weeks' gestation with a hypoplastic left heart.
58	BRAINOT23	This library was constructed using RNA isolated from right temporal lobe tissue removed
,		from a 45-year-old Black male during a brain lobectomy. Pathology for the associated tumor
		tissue indicated dysembryoplastic neuroepithelial tumor of the right temporal lobe. The
		right temporal region dura was consistent with calcifying pseudotumor of the neuraxis. The
		patient presented with convulsive intractable epilepsy, partial epilepsy, and memory
		disturbance. Patient history included obesity, meningitis, backache, unspecified sleep
		apnea, acute stress reaction, acquired knee deformity, and chronic sinusitis. Family
		history included obesity, benign hypertension, cirrhosis of the liver, alcohol abuse,
		hyperlipidemia, cerebrovascular disease, and type II diabetes.

Table 5

	Program	Description	Reference	Parameter Threshold
	ABI FACTURA	A program that removes vector sequences and masks ambiguous bases in nucleic acid sequences.	Perkin-Elmer Applied Biosystems, Foster City, CA.	
	ABI/PARACEL FDF	A Fast Data Finder useful in comparing and annotating amino acid or nucleic acid sequences.	Perkin-Elmer Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA.	Mismatch <50%
	ABI AutoAssembler	A program that assembles nucleic acid sequences.	Perkin-Elmer Applied Biosystems, Foster City, CA.	
74	BLAST	A Basic Local Alignment Search Tool useful in sequence similarity search for amino acid and nucleic acid sequences. BLAST includes five functions: blastp, blastn, blastx, tblastn, and tblastx.	Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410; Altschul, S.F. et al. (1997) Nucleic Acids Res. 25: 3389-3402.	ESTs: Probability value= 1.0E-8 or less Full Length sequences: Probability value= 1.0E-10 or less
	FASTA	A Pearson and Lipman algorithm that searches for similarity between a query sequence and a group of sequences of the same type. FASTA comprises as least five functions: fasta, tfasta, fastx, and ssearch.	Pearson, W.R. and D.J. Lipman (1988) Proc. Natl. Acad Sci. 85:2444-2448; Pearson, W.R. (1990) Methods Enzymol. 183: 63-98; and Smith, T.F. and M. S. Waterman (1981) Adv. Appl. Math. 2:482-489.	ESTs: fasta E value=1.06E-6 Assembled ESTs: fasta Identity= 95% or greater and Match length=200 bases or greater; fastx E value=1.0E-8 or less Full Length sequences: fastx score=100 or greater
	вымрs	A BLocks IMProved Searcher that matches a sequence against those in BLOCKS, PRINTS, DOMO, PRODOM, and PFAM databases to search for gene families, sequence homology, and structural fingerprint regions.	Henikoff, S and J.G. Henikoff, Nucl. Acid Res., 19:6565-72, 1991. J.G. Henikoff and S. Henikoff (1996) Methods Enzymol. 266:88- 105; and Attwood, T.K. et al. (1997) J. Chem. Inf. Comput. Sci. 37: 417-424.	Score=1000 or greater; Ratio of Score/Strength = 0.75 or larger; and, if applicable, Probability value= 1.0E-3 or less
	HMMER	An algorithm for searching a query sequence against hidden Markov model (HMM)-based databases of protein family consensus sequences, such as PFAM.	Krogh, A. et al. (1994) J. Mol. Biol., 235:1501-1531; Sonnhammer, E.L.L. et al. (1988) Nucleic Acids Res. 26:320-322.	Score=10-50 bits for PFAM hits, depending on individual protein families

Table 5 (cont.)

Program	Description	Reference	Parameter Threshold
ProfileScan	An algorithm that searches for structural and sequence motifs in protein sequences that match sequence patterns defined in Prosite.	Gribskov, M. et al. (1988) CABIOS 4:61-66; Gribskov, et al. (1989) Methods Enzymol. 183:146-159; Bairoch, A. et al. (1997) Nucleic Acids Res. 25: 217-221.	Normalized quality score>GCG-specified "HIGH" value for that particular Prosite motif. Generally, score=1.4-2.1.
Phred	A base-calling algorithm that examines automated sequencer traces with high sensitivity and probability.	Ewing, B. et al. (1998) Genome Res. 8:175-185; Ewing, B. and P. Green (1998) Genome Res. 8:186- 194.	
Phrap C	A Phils Revised Assembly Program including SWAT and CrossMatch, programs based on efficient implementation of the Smith-Waterman algorithm, useful in searching sequence homology and assembling DNA sequences.	Smith, T.F. and M. S. Waterman (1981) Adv. Appl. Math. 2:482-489; Smith, T.F. and M. S. Waterman (1981) J. Mol. Biol. 147:195: 197; and Green, P., University of Washington, Seattle, WA.	Score= 120 or greater; Match length= 56 or greater
Consed	A graphical tool for viewing and editing Phrap assemblies	Gordon, D. et al. (1998) Genome Res. 8:195-202.	
SPScan	A weight matrix analysis program that scans protein sequences for the presence of secretory signal peptides.	Nielson, H. et al. (1997) Protein Engineering 10:1-6; Claverie, J.M. and S. Audic (1997) CABIOS 12: 431-439.	Score=3.5 or greater
Motifs	A program that searches amino acid sequences for pattems that matched those defined in Prosite.	Bairoch et al. supra; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI.	÷

What is claimed is:

1. A substantially purified polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-29 and fragments thereof.

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- 2. A substantially purified variant having at least 90% amino acid sequence identity to the amino acid sequence of claim 1.
 - 3. An isolated and purified polynucleotide encoding the polypeptide of claim 1.

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- 4. An isolated and purified polynucleotide variant having at least 90% polynucleotide sequence identity to the polynucleotide of claim 3.
- 5. An isolated and purified polynucleotide which hybridizes under stringent conditions to the polynucleotide of claim 3.
 - 6. An isolated and purified polynucleotide having a sequence which is complementary to the polynucleotide of claim 3.

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- 7. A method for detecting a polynucleotide, the method comprising the steps of:
- (a) hybridizing the polynucleotide of claim 6 to at least one nucleic acid in a sample, thereby forming a hybridization complex; and
- (b) detecting the hybridization complex, wherein the presence of the hybridization complex correlates with the presence of the polynucleotide in the sample.

- 8. The method of claim 7 further comprising amplifying the polynucleotide prior to hybridization.
- 9. An isolated and purified polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:30-58 and fragments thereof.
 - 10. An isolated and purified polynucleotide variant having at least 90% polynucleotide sequence identity to the polynucleotide of claim 9.

11. An isolated and purified polynucleotide having a sequence which is complementary to the polynucleotide of claim 9.

12. An expression vector comprising at least a fragment of the polynucleotide of claim 3.

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- 13. A host cell comprising the expression vector of claim 12.
- 14. A method for producing a polypeptide, the method comprising the steps of:
- a) culturing the host cell of claim 13 under conditions suitable for the expression of the polypeptide; and
 - b) recovering the polypeptide from the host cell culture.
- 15. A pharmaceutical composition comprising the polypeptide of claim 1 in conjunction with a suitable pharmaceutical carrier.

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- 16. A purified antibody which specifically binds to the polypeptide of claim 1.
- 17. A purified agonist of the polypeptide of claim 1.

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18. A purified antagonist of the polypeptide of claim 1.

19. A method for treating or preventing a disorder associated with decreased expression or activity of GTPAP, the method comprising administering to a subject in need of such treatment an effective amount of the pharmaceutical composition of claim 15.

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20. A method for treating or preventing a disorder associated with increased expression or activity of GTPAP, the method comprising administering to a subject in need of such treatment an effective amount of the antagonist of claim 18.

SEQUENCE LISTING

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      HILLMAN, Jennifer L.
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      BANDMAN, Olga
      LAL, Preeti
      YUE, Henry
      LU, Dyung Aina M.
      BAUGHN, Mariah R.
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Arg Asn Ser Ala Thr Cys His Ser Glu Asp Ser Asp Leu Glu Ile

290

<210> 9 <211> 485 <212> PRT

Asp

295

<213> Homo sapiens -

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355

350

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Ser Asp Met His Tyr Arg Val Lys Glu Lys Ile Ile Lys Lys Phe
               365
                                   370
Glu Cys Asn Leu Leu Val Val Cys Ala Asn His Ile Ile Leu Cys
                380
                                   385
Gln Glu Lys Arg Leu Gln Cys Leu Ser Phe Ser Gly Val Lys Glu
                                   400
                395
Arg Glu Trp Gln Met Glu Ser Leu Ile Arg Tyr Ile Lys Val Ile
                                   415
                410
Gly Gly Pro Pro Gly Arg Glu Gly Leu Leu Val Gly Leu Lys Lys
                                   430
                425
Met Tyr Leu Leu Val Tyr Ser Phe Ile Leu Ile Val Lys Asp Tyr
                                   445
Phe Ser Leu Ser Thr Asp Val Leu Gly Asn Leu Thr Trp Lys His
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Val Cys Lys Lys His Tyr Trp Val Phe His Leu Phe Ser Trp Tyr
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Tyr Ile Phe Val Gln
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185
                                   190
Ile Ala Asn Thr Gly Lys Phe Ile Met Thr Ala Ser Ser Asp Thr
               200
                                    205
Thr Val Leu Ile Trp Ser Leu Lys Gly Gln Val Leu Ser Thr Ile
               215
                                    220
Asn Thr Asn Gln Met Asn Asn Thr His Ala Ala Val Ser Pro Cys
                                    235
Gly Arg Phe Val Ala Ser Cys Gly Phe Thr Pro Asp Val Lys Val
                245
Trp Glu Val Cys Phe Gly Lys Lys Gly Glu Phe Gln Glu Val Val
                260
                                    265
Arg Ala Phe Glu Leu Lys Gly His Ser Ala Ala Val His Ser Phe
                                    280
               275
Ala Phe Ser Asn Asp Ser Arg Arg Met Ala Ser Val Ser Lys Asp
                                   .295
               290
Gly Thr Trp Lys Leu Trp Asp Thr Asp Val Glu Tyr Lys Lys
                                    310
               305
Gln Asp Pro Tyr Leu Leu Lys Thr Gly Arg Phe Glu Glu Ala Ala
                                    325
               320
Gly Ala Ala Pro Cys Arg Leu Ala Leu Ser Pro Asn Ala Gln Val
                                    340
Leu Ala Leu Ala Ser Gly Ser Ser Ile His Leu Tyr Asn Thr Arg
Arg Gly Glu Lys Glu Glu Cys Phe Glu Arg Val His Gly Glu Cys
Ile Ala Asn Leu Ser Phe Asp Ile Thr Gly Arg Phe Leu Ala Ser
                380
Cys Gly Asp Arg Ala Val Arg Leu Phe His Asn Thr Pro Gly His
                                    400
                395
Arg Ala Met Val Glu Glu Met Gln Gly His Leu Lys Arg Ala Ser
                                    415
Asn Glu Ser Thr Arg Gln Arg Leu Gln Gln Gln Leu Thr Gln Ala
                                    430
                425
Gln Glu Thr Leu Lys Ser Leu Gly Ala Leu Lys Lys
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                                    10
Ile Asp Val Ala Arg Glu Cys Phe Asn Ile Gly Asn Phe Asn Ser
Leu Met Ala Ile Ile Ser Gly Met Asn Met Ser Pro Val Ser Arg
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40

55

Leu Lys Lys Thr Trp Ala Lys Val Lys Thr Ala Lys Phe Asp Ile

```
Leu Glu His Gln Met Asp Pro Ser Ser Asn Phe Tyr Asn Tyr Arg
                65
Thr Ala Leu Arg Gly Ala Ala Gln Arg Ser Leu Thr Ala His Ser
                                   85
                80
Ser Arg Glu Lys Ile Val Ile Pro Phe Phe Ser Leu Leu Ile Lys
                95
                                   100
Asp Ile Tyr Phe Leu Asn Glu Gly Cys Ala Asn Arg Leu Pro Asn
               110
                                   115
Gly His Val Asn Phe Glu Lys Phe Trp Glu Leu Ala Lys Gln Val
                                   130
               125
Ser Glu Phe Met Thr Trp Lys Gln Val Glu Cys Pro Phe Glu Arg
                                   145
Asp Arg Lys Ile Leu Gln Tyr Leu Leu Thr Val Pro Val Phe Ser
                                   160
               155
Glu Asp Ala Leu Tyr Leu Ala Ser Tyr Glu Ser Glu Gly Pro Glu
Asn His Ile Glu Lys Asp Arg Trp Lys Ser Leu Arg Ser Ser Leu
               185
                                   190
Leu Gly Arg Val
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<211> 694

<212> PRT

<213> Homo sapiens

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<221> misc_feature

<223> Incyte ID No: 1811877CD1

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Leu	Thr	His	Cys	Thr	Leu	Ala	His	Thr	Ala 190	Gly	Val	Val	Leu	Thr 195
Ala	Thr	Ala	Asp		Asn	Leu	Leu	Leu	Tyr 205	Glu	Ala	Arg	Ser	
Arg	Leu	Gln	Lys	Gln	Phe	Ala	Gly	Tyr	Ser 220	Glu	Glu	Val	Leu	
Val	Arg	Phe	Leu		Pro	Glu	Asp	Ser	His	Val	Val	Val	Ala	
Asn	Ser	Pro	Cys		Lys	Val	Phe	Glu	Leu	Gln	Thr	Ser	Ala	Cys
Gln	Ile	Leu	His		His	Thr	Asp	Ile	250 Val	Leu	Ala	Leu	Asp	
Phe	Arg	Lys	Gly		Leu	Phe	Ala	Ser	265 Cys	Ala	Lys	Asp	Gln	
Val	Arg	Ile	Trp		Met	Asn	Lys	Ala	280 Gly	Gln	Val	Met	Cys	
Ala	Gln	Gly	Ser		His	Thr	His	Ser	295 Val	Gly	Thr	Val	Cys	
Ser	Arg	Leu	Lys		Ser	Phe	Leu	Val	310 Thr	Gly	Ser	Gln	Asp	
Thr	Val	Lys	Leu	320 Trp	Pro	Leu	Pro	Lys	325 Ala	Leu	Leu	Ser	Lys	
Thr	Ala	Pro	Asp	335 Asn	Gly	Pro	Ile	Leu	340 Leu	Gln	Ala	Gln	Thr	345 Thr
Gln	Arg	Cys	His	350 Asp	Lys	Asp	Ile	Asn	355 Ser	Val	Ala	Ile	Ala	
Asn	Asp	Lys	Leu	365 Leu	Ala	Thr	Gly	Ser	370 Gln	Asp	Arg	Thr	Ala	
Leu	Trp	Ala	Leu	380 Pro	Gln	Cys	Gln	Leu	385 Leu	Gly	Val	Phe	Ser	390 Gly
His	Arg	Arg	Gly	395 Leu	Trp	Суз	Val	Gln	400 Phe	Ser	Pro	Met	Asp	405 Gln
Val	Leu	Ala	Thr	410 Ala	Ser	Ala	Asp	Gly	415 Thr	Ile	Lys	Leu	Trp	420 Ala
				425					430 Phe				_	435
				440					445 Arg					450
				455					460 Leu					465
				470					475 His					480
				485					490 His					495
				500					505 Asp					510
				515					520 Glu					525
				530					535					540
				545					Lys 550					555
				560					Pro 565					570
				575					Glu 580					585
Glu	Ala	Thr	Met	Leu	Arg	Leu	Arg	Arg	Asp	Gln	Lys	Glu	Ala	Leu

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595
               590
Leu Arg Phe Cys Val Thr Trp Asn Thr Asn Ser Arg His Cys His
               605
                                  610
Glu Ala Gln Ala Val Leu Gly Val Leu Leu Arg Arg Glu Ala Pro
                                  625
               620
Glu Glu Leu Leu Ala Tyr Glu Gly Val Arg Ala Ala Leu Glu Ala
                                   640
               635
Leu Leu Pro Tyr Thr Glu Arg His Phe Gln Arg Leu Ser Arg Thr
                                   655
               650
Leu Gln Ala Ala Ala Phe Leu Asp Phe Leu Trp His Asn Met Lys
               665
                                   670
Leu Pro Val Pro Ala Ala Pro Thr Pro Trp Glu Thr His Lys
                                   685
Gly Ala Leu Pro
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Gly Pro Asp Pro Gln Leu Ala Val Thr Met Gly Phe Thr Gly Phe
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Gly Lys Lys Ala Arg Thr Phe Asp Leu Glu Ala Met Phe Glu Gln
                35
                                   40
Thr Arg Arg Thr Ala Val Glu Arg Ser Arg Lys Thr Leu Glu Ala
                                   55
Arg Glu Lys Glu Glu Glu Met Asn Arg Glu Lys Glu Leu Arg Arg
                                   70
Gln Asn Glu Asp Ile Glu Pro Thr Ser Ser Arg Ser Asn Val Val
Arg Asp Cys Ser Lys Ser Ser Ser Arg Asp Thr Ser Ser Ser Glu
                95
                                  100
Ser Glu Gln Ser Ser Asp Ser Ser Asp Asp Glu Leu Ile Gly Pro
               110
                                  115
Pro Leu Pro Pro Lys Met Val Gly Lys Pro Val Asn Phe Met Glu
                                  130
               125
Glu Asp Ile Leu Gly Pro Leu Pro Pro Pro Leu Asn Glu Glu Glu
               140
                                  145
155
                                  160
Pro Val His Lys Ile Pro Asp Ser His Glu Ile Thr Leu Lys His
               170
                                  175
Gly Thr Lys Thr Val Ser Ala Leu Gly Leu Asp Pro Ser Gly Ala
                                  190 .
               185
Arg Leu Val Thr Gly Gly Tyr Asp Tyr Asp Val Lys Phe Trp Asp
                                  205
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Phe Ala Gly Met Asp Ala Ser Phe Lys Ala Phe Arg Ser Leu Gln

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220
                215
Pro Cys Glu Cys His Gln Ile Lys Ser Leu Gln Tyr Ser Asn Thr
                                    235
                230
Gly Asp Met Ile Leu Val Val Ser Gly Ser Ser Gln Ala Lys Val
                                    250
                245
Ile Asp Arg Asp Gly Phe Glu Val Met Glu Cys Ile Lys Gly Asp
                                    265
Gln Tyr Ile Val Asp Met Ala Asn Thr Lys Gly His Thr Ala Met
                                    280
Leu His Thr Gly Ser Trp His Pro Lys Ile Lys Gly Glu Phe Met
                290
Thr Cys Ser Asn Asp Ala Thr Val Arg Thr Trp Glu Val Glu Asn
                                    310
                305
Pro Lys Lys Gln Lys Ser Val Phe Lys Pro Arg Thr Met Gln Gly
                                    325
                320
Lys Lys Val Ile Pro Thr Thr Cys Thr Tyr Ser Arg Asp Gly Asn
                                    340
                335
Leu Ile Ala Ala Cys Gln Asn Gly Ser Ile Gln Ile Trp Asp
                                    355
                350
Arg Asn Leu Thr Val His Pro Lys Phe His Tyr Lys Gln Ala His
                                    370
                365
Asp Ser Gly Thr Asp Thr Ser Cys Val Thr Phe Ser Tyr Asp Gly
                                    385
Asn Val Leu Ala Ser Arg Gly Gly Asp Asp Ser Leu Lys Leu Trp
                395
                                    400
Asp Ile Arg Gln Phe Asn Lys Pro Leu Phe Ser Ala Ser Gly Leu
                                    415
Pro Thr Met Phe Pro Met Thr Asp Cys Cys Phe Ser Pro Asp Asp
                425
Lys Leu Ile Val Thr Gly Thr Ser Ile Gln Arg Gly Cys Gly Ser
                                    445
                440
Gly Lys Leu Val Phe Phe Glu Arg Arg Thr Phe Gln Arg Val Tyr
                455
                                    460
Glu Ile Asp Ile Thr Asp Ala Ser Val Val Arg Cys Leu Trp His
                                    475
                470
Pro Lys Leu Asn Gln Ile Met Val Gly Thr Gly Asn Gly Leu Ala
                                    490
                485
Lys Val Tyr Tyr Asp Pro Asn Lys Ser Gln Arg Gly Ala Lys Leu
                                    505
                500
Cys Val Val Lys Thr Gln Arg Lys Ala Lys Gln Ala Glu Thr Leu
                                    520
                515
Thr Gln Asp Tyr Ile Ile Thr Pro His Ala Leu Pro Met Phe Arg
                                    535
Glu Pro Arg Gln Arg Ser Thr Arg Lys Gln Leu Glu Lys Asp Arg
Leu Asp Pro Leu Lys Ser His Lys Pro Glu Pro Pro Val Ala Gly
Pro Gly Arg Gly Gly Arg Val Gly Thr His Gly Gly Thr Leu Ser
                575
Ser Tyr Ile Val Lys Asn Ile Ala Leu Asp Lys Thr Asp Asp Ser
                                    595
Asn Pro Arg Glu Ala Ile Leu Arg His Ala Lys Ala Ala Glu Asp
                605
                                   610
Ser Pro Tyr Trp Val Ser Pro Ala Tyr Ser Lys Thr Gln Pro Lys
                                 625
                                                         630
                620
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Thr Met Phe Ala Gln Val Glu Ser Asp Asp Glu Glu Ala Lys Asn

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Glu Pro Glu Trp Lys Lys Arg Lys Ile
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Arg Ile Lys Val Ile Ser Met Gly Asn Ala Glu Val Gly Lys Ser
Cys Ile Ile Lys Arg Tyr Cys Glu Lys Arg Phe Val Ser Lys Tyr
                35
                                    40
Leu Ala Thr Ile Gly Ile Asp Tyr Gly Val Thr Lys Val His Val
                 50
Arg Asp Arg Glu Ile Lys Val Asn Ile Phe Asp Met Ala Gly His
                                    70
                65
Pro Phe Phe Tyr Glu Val Arg Asn Glu Phe Tyr Lys Asp Thr Gln
                80
                                    85
Gly Val Ile Leu Val Tyr Asp Val Gly Gln Lys Asp Ser Phe Asp
                                    100
                 95
Ala Leu Asp Ala Trp Leu Ala Glu Met Lys Gln Glu Leu Gly Pro
                                    115
               110
His Gly Asn Met Glu Asn Ile Ile Phe Val Val Cys Ala Asn Lys
                                   130
Ile Asp Cys Thr Lys His Arg Cys Val Asp Glu Ser Glu Gly Arg
Leu Trp Ala Glu Ser Lys Gly Phe Leu Tyr Phe Glu Thr Ser Ala
                                    160
Gln Thr Gly Glu Gly Ile Asn Glu Met Phe Gln Ile His Leu Gly
                                   175
                                                        180
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                                    10
Pro Arg His Pro Val Cys Leu Leu Val Leu Gly Met Ala Gly Ser
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Gly Lys Thr Thr Phe Val Gln Arg Leu Thr Gly His Leu His Ala
                 35
                                     40
Gln Gly Thr Pro Pro Tyr Val Ile Asn Leu Asp Pro Ala Val His
                                     55
Glu Val Pro Phe Pro Ala Asn Ile Asp Ile Arg Asp Thr Val Lys
Tyr Lys Glu Val Met Lys Gln Tyr Gly Leu Gly Pro Asn Gly Gly
                 80
Ile Val Thr Ser Leu Asn Leu Phe Ala Thr Arg Phe Asp Gln Val
                                    100
                 95
Met Lys Phe Ile Glu Lys Ala Gln Asn Met Ser Lys Tyr Val Leu
                                    115
                110
Ile Asp Thr Pro Gly Gln Ile Glu Val Phe Thr Trp Ser Ala Ser
                                    130
                125
Gly Thr Ile Ile Thr Glu Ala Leu Ala Ser Ser Phe Pro Thr Val
                                    145
                140
Val Ile Tyr Val Met Asp Thr Ser Arg Ser Thr Asn Pro Val Thr
                                    160
                155
Phe Met Ser Asn Met Leu Tyr Ala Cys Ser Ile Leu Tyr Lys Thr
                                    175
                170
Lys Leu Pro Phe Ile Val Val Met Asn Lys Thr Asp Ile Ile Asp
                                    190
His Ser Phe Ala Val Glu Trp Met Gln Asp Phe Glu Ala Phe Gln
                                    205
Asp Ala Leu Asn Gln Glu Thr Thr Tyr Val Ser Asn Leu Thr Arg
                                    220
Ser Met Ser Leu Val Leu Asp Glu Phe Tyr Ser Ser Leu Arg Val
                                    235
                230
Val Gly Val Ser Ala Val Leu Gly Thr Gly Leu Asp Glu Leu Phe
                                    250
Val Gln Val Thr Ser Ala Ala Glu Glu Tyr Glu Arg Glu Tyr Arg
                                    265
                260
Pro Glu Tyr Glu Arg Leu Lys Lys Ser Leu Ala Asn Ala Glu Ser
                                    280
                275
Gln Gln Gln Arg Glu Gln Leu Glu Arg Leu Arg Lys Asp Met Gly
                290
                                    295
Ser Val Ala Leu Asp Ala Gly Thr Ala Lys Asp Ser Leu Ser Pro
                                    310
                305
Val Leu His Pro Ser Asp Leu Ile Leu Thr Arg Gly Thr Leu Asp
                                    325
Glu Glu Asp Glu Glu Ala Asp Ser Asp Thr Asp Asp Ile Asp His
Arg Val Thr Glu Glu Ser His Glu Glu Pro Ala Phe Gln Asn Phe
                                    355
Met Gln Glu Ser Met Ala Gln Tyr Trp Lys Arg Asn Asn Lys
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<220>

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380 -
                                     385
 Gly Pro Glu Asn Pro Gly Leu Gly Leu Leu Asn Asp Ile Leu Leu
                                     400
                 395
 Thr Tyr Cys Met Tyr His Phe Asp Leu Gly Tyr Val Gln Gly Met
 Ser Asp Leu Leu Ser Pro Ile Leu Tyr Val Ile Gln Asn Glu Val
                                     430
 Asp Ala Phe Trp Cys Phe Cys Gly Phe Met Glu Leu Val Gln Gly
                 440
 Asn Phe Glu Glu Ser Gln Glu Thr Met Lys Arg Gln Leu Gly Arg
                                     460
 Leu Leu Leu Leu Arg Val Leu Asp Pro Leu Leu Cys Asp Phe
                 470
                                     475
 Leu Asp Ser Gln Asp Ser Gly Ser Leu Cys Phe Cys Phe Arg Trp
                                     490
 Leu Leu Ile Trp Phe Lys Arg Glu Phe Pro Phe Pro Asp Val Leu
                                     505
                 500
 Arg Leu Trp Glu Val Leu Trp Thr Gly Leu Pro Gly Pro Asn Leu
                                     520
                 515
 His Leu Leu Val Ala Cys Ala Ile Leu Asp Met Glu Arg Asp Thr
                                     535
 Leu Met Leu Ser Gly Phe Gly Ser Asn Glu Ile Leu Lys His Ile
                 545
 Asn Glu Leu Thr Met Lys Leu Ser Val Glu Asp Val Leu Thr Arg
 Ala Glu Ala Leu His Arg Gln Leu Thr Ala Cys Thr Arg Ala Ala
                 575
 Pro Gln Arg Ala Gly Asp Pro Gly Ala Gly Pro Ala Thr Gln Ser
                                     595
                 590
. Pro Thr Ala Pro Arg Pro Pro Pro Pro Arg Cys Leu Cys Thr Pro
                                     610
                 605
 Thr Arg Ala Pro Pro Thr Pro Pro Pro Ser Thr Asp Thr Ala Pro
                                     625
                 620
 Gln Pro Asp Ser Ser Leu Glu Ile Leu Pro Glu Glu Glu Asp Glu
                                     640
 Gly Ala Asp Ser
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 <211> 698
 <212> PRT
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 <221> misc feature
 <223> Incyte ID No: 2707969CD1
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Ala Glu Lys Ala Ile Glu Leu Arg Leu Ala Lys Ile Asp His Thr

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Ala Ile His Pro His Leu Leu Asp Met Lys Ile Gly Gln Gly Lys
                65
                                   70
Tyr Glu Pro Gly Phe Phe Pro Lys Leu Gln Ser Asp Val Leu Ser
                                    85
                 80
Thr Gly Pro Ala Ser Asn Lys Trp Thr Lys Arg Asn Ala Pro Ala
                 95
                                   100
Gln Trp Arg Arg Lys Asp Arg Gln Lys Gln His Thr Glu His Leu
                                   115
                110
Arg Leu Asp Asn Asp Gln Arg Glu Lys Tyr Ile Gln Glu Ala Arg
                                   130
                125 ,
Thr Met Gly Ser Thr Ile Arg Gln Pro Lys Leu Ser Asn Leu Ser
Pro Ser Val Ile Ala Gln Thr Asn Trp Lys Phe Val Glu Gly Leu
                                    160
Leu Lys Glu Cys Arg Asn Lys Thr Lys Arg Met Leu Val Glu Lys
                                   175
                170
Met Gly Arg Glu Ala Val Glu Leu Gly His Gly Glu Val Asn Ile
                                   190
                185
Thr Gly Val Glu Glu Asn Thr Leu Ile Ala Ser Leu Cys Asp Leu
                200
                                   205
Leu Glu Arg Ile Trp Ser His Gly Leu Gln Val Lys Gln Gly Lys
                215
                                   220
Ser Ala Leu Trp Ser His Leu Leu His Tyr Gln Asp Asn Arg Gln
                230
                                   235
Arg Lys Leu Thr Ser Gly Ser Leu Ser Thr Ser Gly Ile Leu Leu
                                   250
                245
Asp Ser Glu Arg Arg Lys Ser Asp Ala Ser Ser Leu Met Pro Pro
                                   265
                260
Leu Arg Ile Ser Leu Ile Gln Asp Met Arg His Ile Gln Asn Ile
                                   280
                275
Gly Glu Ile Lys Thr Asp Val Gly Lys Ala Arg Ala Trp Val Arg
                                   295
Leu Ser Met Glu Lys Lys Leu Leu Ser Arg His Leu Lys Gln Leu
                                   310
                305
Leu Ser Asp His Glu Leu Thr Lys Lys Leu Tyr Lys Arg Tyr Ala
                                   325
Phe Leu Arg Cys Asp Asp Glu Lys Glu Gln Phe Leu Tyr His Leu
                335
                                   340
Leu Ser Phe Asn Ala Val Asp Tyr Phe Cys Phe Thr Asn Val Phe
               350
                                   355
Thr Thr Ile Leu Ile Pro Tyr His Ile Leu Ile Val Pro Ser Lys
               365
                                   370
Lys Leu Gly Gly Ser Met Phe Thr Ala Asn Pro Trp Ile Cys Ile
                                   385
                380
Ser Gly Glu Leu Gly Glu Thr Gln Ile Met Gln Ile Pro Arg Asn
                395
                                   400
Val Leu Glu Met Thr Phe Glu Cys Gln Asn Leu Gly Lys Leu Thr
                                   415
                410 .
Thr Val Gln Ile Gly His Asp Asn Ser Gly Leu Tyr Ala Lys Trp
                425
                                   430
Leu Val Glu Tyr Val Met Val Arg Asn Glu Ile Thr Gly His Thr
                440 .
                                   445
Tyr Lys Phe Pro Cys Gly Arg Trp Leu Gly Lys Gly Met Asp Asp
                                   460
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Gly Ser Leu Glu Arg Ile Leu Val Gly Glu Leu Leu Thr Ser Gln
                470
                                    475
Pro Glu Val Asp Glu Arg Pro Cys Arg Thr Pro Pro Leu Gln Gln
                                    490
                485
Ser Pro Ser Val Ile Arg Arg Leu Val Thr Ile Ser Pro Asn Asn
                                    505
Lys Pro Lys Leu Asn Thr Gly Gln Ile Gln Glu Ser Ile Gly Glu
                                    520
Ala Val Asn Gly Ile Val Lys His Phe His Lys Pro Glu Lys Glu
                530
Arg Gly Ser Leu Thr Leu Leu Leu Cys Gly Glu Cys Gly Leu Val
                                    550
                545
Ser Ala Leu Glu Gln Ala Phe Gln His Gly Phe Lys Ser Pro Arg
                                    565
                560
Leu Phe Lys Asn Val Phe Ile Trp Asp Phe Leu Glu Lys Ala Gln
                                    580
                575
Thr Tyr Tyr Glu Thr Leu Glu Lys Asn Glu Val Val Pro Glu Glu
                                   595
                590
Asn Trp His Thr Arg Ala Arg Asn Phe Cys Arg Phe Val Thr Ala
                605
                                    610
Ile Asn Asn Thr Pro Arg Asn Ile Gly Lys Asp Gly Lys Phe Gln
                                    625
Met Leu Val Cys Leu Gly Ala Arg Asp His Leu Leu His His Trp
                                    640
Ile Ala Leu Leu Ala Asp Cys Pro Ile Thr Ala His Met Tyr Glu
                                    655
                650
Asp Val Ala Leu Ile Lys Asp His Thr Leu Val Asn Ser Leu Ile
                                    670
                665
Arg Val Leu Gln Thr Leu Gln Glu Phe Asn Ile Thr Leu Glu Thr
                                    685
                680
Ser Leu Val Lys Gly Ile Asp Ile
                695
<210> 18
<211> 396
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<213> Homo sapiens
<220>
<221> misc feature
<223> Incyte ID No: 2817769CD1
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Met Pro Pro Lys Lys Gly Gly Asp Gly Ile Lys Pro Pro Pro Ile
                                     10
Ile Gly Arg Phe Gly Thr Ser Leu Lys Ile Gly Ile Val Gly Leu
Pro Asn Val Gly Lys Ser Thr Phe Phe Asn Val Leu Thr Asn Ser
                                     40
                 35
Gln Ala Ser Ala Glu Asn Phe Pro Phe Cys Thr Ile Asp Pro Asn
                                     55
Glu Ser Arg Val Pro Val Pro Asp Glu Arg Phe Asp Phe Leu Cys
                                     70
                 65
Gln Tyr His Lys Pro Ala Ser Lys Ile Pro Ala Phe Leu Asn Val
                                                         90
                 80
                                     85
```

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```
Val Asp Ile Ala Gly Leu Val Lys Gly Ala His Asn Gly Gln Gly
                                   100
                95
Leu Gly Asn Ala Phe Leu Ser His Ile Ser Ala Cys Asp Gly Ile
                                   115
Phe His Leu Thr Arg Ala Phe Glu Asp Asp Ile Thr His Val
                                  130
               125
Glu Gly Ser Val Asp Pro Ile Arg Asp Ile Glu Ile Ile His Glu
               140
                                   145
Glu Leu Gln Leu Lys Asp Glu Glu Met Ile Gly Pro Ile Ile Asp
               155
                                   160
Lys Leu Glu Lys Val Ala Val Arg Gly Gly Asp Lys Leu Lys
                                   175
               170
Pro Glu Tyr Asp Ile Met Cys Lys Val Lys Ser Trp Val Ile Asp
                                   190
Gln Lys Lys Pro Val Arg Phe Tyr His Asp Trp Asn Asp Lys Glu
                                   205
Ile Glu Val Leu Asn Lys His Leu Phe Leu Thr Ser Lys Pro Met
                                   220
Val Tyr Leu Val Asn Leu Ser Glu Lys Asp Tyr Ile Arg Lys Lys
                                   235
               230
Asn Lys Trp Leu Ile Lys Ile Lys Glu Trp Val Asp Lys Tyr Asp
                                   250
               245
Pro Gly Ala Leu Val Ile Pro Phe Ser Gly Ala Leu Glu Leu Lys
                                   265
               260
Leu Gln Glu Leu Ser Ala Glu Glu Arg Gln Lys Tyr Leu Glu Ala
                                   280
               275
Asn Met Thr Gln Ser Ala Leu Pro Lys Ile Ile Lys Ala Gly Phe
                                   295
               290
Ala Ala Leu Gln Leu Glu Tyr Phe Phe Thr Ala Gly Pro Asp Glu
                                   310
               305
Val Arg Ala Trp Thr Ile Arg Lys Gly Thr Lys Ala Pro Gln Ala
                                   325
                320
Ala Gly Lys Ile His Thr Asp Phe Glu Lys Gly Phe Ile Met Ala
                                   340
                335
Glu Val Met Lys Tyr Glu Asp Phe Lys Glu Glu Gly Ser Glu Asn
                                    355
                350
Ala Val Lys Ala Ala Gly Lys Tyr Arg Gln Gln Gly Arg Asn Tyr
                                    370
Ile Val Glu Asp Gly Asp Ile Ile Phe Phe Lys Phe Asn Thr Pro
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                380
Gln Gln Pro Lys Lys Lys
                395
<210> 19
<211> 634
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<210> 19
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<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No: 2917557CD1

<400> 19
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Glu Glu Arg Ala Tyr Asp Lys Ala Lys Arg Arg Ile Glu Lys Arg
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Arg Leu Glu His Ser Lys Asn Val Asn Thr Glu Lys Leu Arg Ala
Pro Ile Ile Cys Val Leu Gly His Val Asp Thr Gly Lys Thr Lys
Ile Leu Asp Lys Leu Arg His Thr His Val Gln Asp Gly Glu Ala
                 65
Gly Gly Ile Thr Gln Gln Ile Gly Ala Thr Asn Val Pro Leu Glu
                                     85
Ala Ile Asn Glu Gln Thr Lys Met Ile Lys Asn Phe Asp Arg Glu
                                    100
                 95
Asn Val Arg Ile Pro Gly Met Leu Ile Ile Asp Thr Pro Gly His
                                    115
Glu Ser Phe Ser Asn Leu Arg Asn Arg Gly Ser Ser Leu Cys Asp
                                    130
                125
Ile Ala Ile Leu Val Val Asp Ile Met His Gly Leu Glu Pro Gln
                                    145
                140
Thr Ile Glu Ser Ile Asn Leu Leu Lys Ser Lys Lys Cys Pro Phe
                                    160
Ile Val Ala Leu Asn Lys Ile Asp Arg Leu Tyr Asp Trp Lys Lys
                170
Ser Pro Asp Ser Asp Val Ala Ala Thr Leu Lys Lys Gln Lys Lys
Asn Thr Lys Asp Glu Phe Glu Glu Arg Ala Lys Ala Ile Ile Val
                200
Glu Phe Ala Gln Gln Gly Leu Asn Ala Ala Leu Phe Tyr Glu Asn
                                    220
                215
Lys Asp Pro Arg Thr Phe Val Ser Leu Val Pro Thr Ser Ala His
                                    235
                230
Thr Gly Asp Gly Met Gly Ser Leu Ile Tyr Leu Leu Val Glu Leu
                                    250
                245
Thr Gln Thr Met Leu Ser Lys Arg Leu Ala His Cys Glu Glu Leu
                                    265
                260
Arg Ala Gln Val Met Glu Val Lys Ala Leu Pro Gly Met Gly Thr
                                    280
                275
Thr Ile Asp Val Ile Leu Ile Asn Gly Arg Leu Lys Glu Gly Asp
                                    295
                290
Thr Ile Ile Val Pro Gly Val Glu Gly Pro Ile Val Thr Gln Ile
                305
                                    310
Arg Gly Leu Leu Pro Pro Pro Met Lys Glu Leu Arg Val Lys
Asn Gln Tyr Glu Lys His Lys Glu Val Glu Ala Ala Gln Gly Val
Lys Ile Leu Gly Lys Asp Leu Glu Lys Thr Leu Ala Gly Leu Pro
                350
Leu Leu Val Ala Tyr Lys Glu Asp Glu Ile Pro Val Leu Lys Asp
                                    370
                                                         375
Glu Leu Ile His Glu Leu Lys Gln Thr Leu Asn Ala Ile Lys Leu
                                    385
                380
Glu Glu Lys Gly Val Tyr Val Gln Ala Ser Thr Leu Gly Ser Leu
                                    400
                395
Glu Ala Leu Leu Glu Phe Leu Lys Thr Ser Glu Val Pro Tyr Ala
                                                         420
                                    415
                410
```

```
Gly Ile Asn Ile Gly Pro Val His Lys Lys Asp Val Met Lys Ala
               425
                                  430
Ser Val Met Leu Glu His Asp Pro Gln Tyr Ala Val Ile Leu Ala
                                    445
                440
Phe Asp Val Arg Ile Glu Arg Asp Ala Gln Glu Met Ala Asp Ser
                455
                                   460
Leu Gly Val Arg Ile Phe Ser Ala Glu Ile Ile Tyr His Leu Phe
                                   475
                470
Asp Ala Phe Thr Lys Tyr Arg Gln Asp Tyr Lys Lys Gln Lys Gln
                                   490
                485
Glu Glu Phe Lys His Ile Ala Val Phe Pro Cys Lys Ile Lys Ile
                                    505
Leu Pro Gln Tyr Ile Phe Asn Ser Arg Asp Pro Ile Val Met Gly
                                    520
                515
Val Thr Val Glu Ala Gly Gln Val Lys Gln Gly Thr Pro Met Cys
                                    535
Val Pro Ser Lys Asn Phe Val Asp Ile Gly Ile Val Thr Ser Ile
                                    550
                545
Glu Ile Asn His Lys Gln Val Asp Val Ala Lys Lys Gly Gln Glu
                560
                                    565
Val Cys Val Lys Ile Glu Pro Ile Pro Gly Glu Ser Pro Lys Met
                575
                                   580
Phe Gly Arg His Phe Glu Ala Thr Asp Ile Leu Val Ser Lys Ile
                                   595
                590
Ser Arg Gln Ser Ile Asp Ala Leu Lys Asp Trp Phe Arg Asp Glu
               605
                                   610
Met Gln Lys Ser Asp Trp Gln Leu Ile Val Glu Leu Lys Lys Val
                                    625
Phe Glu Ile Ile
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<210> 20 <211> 196 <212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 3421335CD1

<400> 20

Met Gly Ser Val Asn Ser Arg Gly His Lys Ala Glu Ala Gln Val 10 Val Met Met Gly Leu Asp Ser Ala Gly Lys Thr Thr Leu Leu Tyr 20 25 Lys Leu Lys Gly His Gln Leu Val Glu Thr Leu Pro Thr Val Gly 35 40 Phe Asn Val Glu Pro Leu Lys Ala Pro Gly His Val Ser Leu Thr 50 Leu Trp Asp Val Gly Gln Ala Pro Leu Arg Ala Ser Trp Lys 70 65 Asp Tyr Leu Glu Gly Thr Asp Ile Leu Val Tyr Val Leu Asp Ser 80 -85 Thr Asp Glu Ala Arg Leu Pro Glu Ser Ala Ala Glu Leu Thr Glu 100

```
        Val
        Leu
        Asn
        Asp
        Pro
        Asn
        Met
        Ala
        Gly
        Val
        Pro
        Phe
        Leu
        Leu
        Val
        Leu
        Phe
        Leu
        Phe
        Leu
        Phe
        Leu
        Leu
        Leu
        Leu
        Leu
        Phe
        Asp
        Ala
        Leu
        Leu
        Leu
        Phe
        Gln
        Asp
        His
        Cys
        Trp
        Glu
        Ala
        Leu
        Thr
        Gly
        Glu
        Gly
        Leu
        Pho
        Glu
        Asp
        His
        Cys
        Trp
        Glu
        Ala
        Ala
        Leu
        Thr
        Gly
        Glu
        Gly
        Leu
        Pho
        Glu
        Ala
        Gly
        Ala
        Ala
        Ala
        Leu
        Thr
        Gly
        Glu
        Gly
        Leu
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        Leu
        Leu
        Leu
        Leu
        Leu
        Ala
        Ala</th
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<210> 21 <211> 446 <212> PRT <213> Homo sapiens

<220>
<221> misc_feature

<223> Incyte ID No: 605761CD1

<400> 21

Met Ala Ala Arg Lys Gly Arg Arg Arg Thr Cys Glu Thr Gly Glu Pro Met Glu Ala Glu Ser Gly Asp Thr Ser Ser Glu Gly Pro Ala Gln Val Tyr Leu Pro Gly Arg Gly Pro Pro Leu Arg Glu Gly Glu Glu Leu Val Met Asp Glu Glu Ala Tyr Val Leu Tyr His Arg Ala 55 50 Gln Thr Gly Ala Pro Cys Leu Ser Phe Asp Ile Val Arg Asp His 65 Leu Gly Asp Asn Arg Thr Glu Leu Pro Leu Thr Leu Tyr Leu Cys 85 80 Ala Gly Thr Gln Ala Glu Ser Ala Gln Ser Asn Arg Leu Met Met 95 100 Leu Arg Met His Asn Leu His Gly Thr Lys Pro Pro Pro Ser Glu 110 115 Gly Ser Asp Glu Glu Glu Glu Glu Glu Asp Glu Glu Asp Glu Glu 130 125 Glu Arg Lys Pro Gln Leu Glu Leu Ala Met Val Pro His Tyr Gly 145 Gly Ile Asn Arg Val Arg Val Ser Trp Leu Gly Glu Glu Pro Val 160 155 Ala Gly Val Trp Ser Glu Lys Gly Gln Val Glu Val Phe Ala Leu Arg Arg Leu Leu Gln Val Val Glu Glu Pro Gln Ala Leu Ala Ala 190 185 Phe Leu Arg Asp Glu Gln Ala Gln Met Lys Pro Ile Phe Ser Phe 205 Ala Gly His Met Gly Glu Gly Phe Ala Leu Asp Trp Ser Pro Arg 215 220

```
Val Thr Gly Arg Leu Leu Thr Gly Asp Cys Gln Lys Asn Ile His
                                  235
               230
Leu Trp Thr Pro Thr Asp Gly Gly Ser Trp His Val Asp Gln Arg
                                   250
               245
Pro Phe Val Gly His Thr Arg Ser Val Glu Asp Leu Gln Trp Ser
                                   265
               260
Pro Thr Glu Asn Thr Val Phe Ala Ser Cys Ser Ala Asp Ala Ser
                                   280
               275
Ile Arg Ile Trp Asp Ile Arg Ala Ala Pro Ser Lys Ala Cys Met
                                   295
               290
Leu Thr Thr Ala Thr Ala His Asp Gly Asp Val Asn Val Ile Ser
                                   310
               305
Trp Ser Arg Arg Glu Pro Phe Leu Leu Ser Gly Gly Asp Asp Gly
                                   325
Ala Leu Lys Ile Trp Asp Leu Arg Gln Phe Lys Ser Gly Ser Pro
                                   340
Val Ala Thr Phe Lys Gln His Val Ala Pro Val Thr Ser Val Glu
                                   355
               350
Trp His Pro Gln Asp Ser Gly Val Phe Ala Ala Ser Gly Ala Asp
               365
                                   370
His Gln Ile Thr Gln Trp Asp Leu Ala Val Glu Arg Asp Pro Glu
               380
                                   385
Ala Gly Asp Val Glu Ala Asp Pro Gly Leu Ala Asp Leu Pro Gln
                                   400
               395
Gln Leu Leu Phe Val His Gln Gly Glu Thr Glu Leu Lys Glu Leu
               410
                                   415
His Trp His Pro Gln Cys Pro Gly Leu Leu Val Ser Thr Ala Leu
                                   430
               425
Ser Gly Phe Thr Ile Phe Arg Thr Ile Ser Val
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<210> 22
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<211> 265

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 483862CD1

<400> 22

Met Ser Ser Gly Leu Arg Ala Ala Asp Phe Pro Arg Trp Lys Arg 10 His Ile Ser Glu Gln Leu Arg Arg Arg Asp Arg Leu Gln Arg Gln 20 25 Ala Phe Glu Glu Ile Ile Leu Gln Tyr Asn Lys Leu Leu Glu Lys 40 . 35 Ser Asp Leu His Ser Val Leu Ala Gln Lys Leu Gln Ala Glu Lys 55 50 His Asp Val Pro Asn Arg His Glu Ile Ser Pro Gly His Asp Gly 70 65 Thr Trp Asn Asp Asn Gln Leu Gln Glu Met Ala Gln Leu Arg Ile 85 80 Lys His Gln Glu Glu Leu Thr Glu Leu His Lys Lys Arg Gly Glu

```
95 -
                                    100
Leu Ala Gln Leu Val Ile Asp Leu Asn Asn Gln Met Gln Arg Lys
                                    115
                110
Asp Arg Glu Met Gln Met Asn Glu Ala Lys Ile Ala Glu Cys Leu
                                    130
Gln Thr Ile Ser Asp Leu Glu Thr Glu Cys Leu Asp Leu Arg Thr
Lys Leu Cys Asp Leu Glu Arg Ala Asn Gln Thr Leu Lys Asp Glu
                                    160
Tyr Asp Ala Leu Gln Ile Thr Phe Thr Ala Leu Glu Gly Lys Leu
                                    175
                170
Arg Lys Thr Thr Glu Glu Asn Gln Glu Leu Val Thr Arg Trp Met
                                    190
                185
Ala Glu Lys Ala Gln Glu Ala Asn Arg Leu Asn Ala Glu Asn Glu
                                   .205
                200
Lys Asp Ser Arg Arg Arg Gln Ala Arg Leu Gln Lys Glu Leu Ala
                                    220
                215
Glu Ala Ala Lys Glu Pro Leu Pro Val Glu Gln Asp Asp Asp Ile
                                    235
                230
Glu Val Ile Val Asp Glu Thr Ser Asp His Thr Glu Glu Thr Ser
                                    250
                245
Pro Val Arg Ala Ile Ser Arg Ala Ala Thr
                                    265
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<210> 23 <211> 185 <212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 1256777CD1

<400> 23

Met Leu Lys Ala Lys Ile Leu Phe Val Gly Pro Cys Glu Ser Gly 10 Lys Thr Val Leu Ala Asn Phe Leu Thr Glu Ser Ser Asp Ile Thr 20 Glu Tyr Ser Pro Thr Gln Gly Val Arg Ile Leu Glu Phe Glu Asn 40 Pro His Val Thr Ser Asn Asn Lys Gly Thr Gly Cys Glu Phe Glu 55 Leu Trp Asp Cys Gly Gly Asp Ala Lys Phe Glu Ser Cys Trp Pro 70 Ala Leu Met Lys Asp Ala His Gly Val Val Ile Val Phe Asn Ala Asp Ile Pro Ser His Arg Lys Glu Met Glu Met Trp Tyr Ser Cys 100 95 Phe Val Gln Gln Pro Ser Leu Gln Asp Thr Gln Cys Met Leu Ile 115 Ala His His Lys Pro Gly Ser Gly Asp Asp Lys Gly Ser Leu Ser 130 125 Leu Ser Pro Pro Leu Asn Lys Leu Lys Leu Val His Ser Asn Leu 150 145 140

```
Glu Asp Asp Pro Glu Glu Ile Arg Met Glu Phe Ile Lys Tyr Leu
                                   160
               155
Lys Ser Ile Ile Asn Ser Met Ser Glu Ser Arg Asp Arg Glu Glu
                                   175
                170
Met Ser Ile Met Thr
                185
<210> 24
<211> 554
<212> PRT
<213> Homo sapiens
<220>
<221> misc feature
<223> Incyte ID No: 2198779CD1
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                                     10
Pro Ser Glu Gly Leu Pro Arg Arg Gly Ala Gly Leu Arg Arg Ser
                 20
Glu Glu Glu Glu Glu Asp Glu Asp Val Asp Leu Ala Gln Val
                                     40
                 35
Leu Ala Tyr Leu Leu Arg Arg Gly Gln Val Arg Leu Val Gln Gly
                 50
                                     55
Gly Gly Ala Ala Asn Leu Gln Phe Ile Gln Ala Leu Leu Asp Ser
                                     70
                 65
Glu Glu Glu Asn Asp Arg Ala Trp Asp Gly Arg Leu Gly Asp Arg
                                     85
                 80
Tyr Asn Pro Pro Val Asp Ala Thr Pro Asp Thr Arg Glu Leu Glu
                                   100
Phe Asn Glu Ile Lys Thr Gln Val Glu Leu Ala Thr Gly Gln Leu
                                    115
                110
Gly Leu Arg Arg Ala Ala Gln Lys His Ser Phe Pro Arg Met Leu
                                    130
His Gln Arg Glu Arg Gly Leu Cys His Arg Gly Ser Phe Ser Leu
                                    145
                140
Gly Glu Gln Ser Arg Val Ile Ser His Phe Leu Pro Asn Asp Leu
                                   160
                155
Gly Phe Thr Asp Ser Tyr Ser Gln Lys Ala Phe Cys Gly Ile Tyr
                                   175
                170
Ser Lys Asp Gly Gln Ile Phe Met Ser Ala Cys Gln Asp Gln Thr
                                   190
                185
Ile Arg Leu Tyr Asp Cys Arg Tyr Gly Arg Phe Arg Lys Phe Lys
                                    205
                200
Ser Ile Lys Ala Arg Asp Val Gly Trp Ser Val Leu Asp Val Ala
                215
                                    220
Phe Thr Pro Asp Gly Asn His Phe Leu Tyr Ser Ser Trp Ser Asp
                230
                                    235
                                                        240
Tyr Ile His Ile Cys Asn Ile Tyr Gly Glu Gly Asp Thr His Thr
                245
                                    250
Ala Leu Asp Leu Arg Pro Asp Glu Arg Arg Phe Ala Val Phe Ser
                260
                                    265
Ile Ala Val Ser Ser Asp Gly Arg Glu Val Leu Gly Gly Ala Asn
                                    280
                275
```

```
Asp Gly Cys Leu Tyr Val Phe Asp Arg Glu Gln Asn Arg Arg Thr
                                    295 '
Leu Gln Ile Glu Ser His Glu Asp Asp Val Asn Ala Val Ala Phe
                                    310
                305
Ala Asp Ile Ser Ser Gln Ile Leu Phe Ser Gly Gly Asp Asp Ala
                                    325
                320
Ile Cys Lys Val Trp Asp Arg Arg Thr Met Arg Glu Asp Asp Pro
Lys Pro Val Gly Ala Leu Ala Gly His Gln Asp Gly Ile Thr Phe
                                    355
Ile Asp Ser Lys Gly Asp Ala Arg Tyr Leu Ile Ser Asn Ser Lys
                                    370
                365
Asp Gln Thr Ile Lys Leu Trp Asp Ile Arg Arg Phe Ser Ser Arg
                380
                                    385
Glu Gly Met Glu Ala Ser Arg Gln Ala Ala Thr Gln Gln Asn Trp
                                    400
                395
Asp Tyr Arg Trp Gln Gln Val Pro Lys Lys Gly Phe Thr Leu His
                                    415
                410
Pro Tyr Pro Ala Trp Arg Lys Leu Lys Leu Pro Gly Asp Ser Ser
                                    430
                425
Leu Met Thr Tyr Arg Gly His Gly Val Leu His Thr Leu Ile Arg
                                    445
                440
Cys Arg Phe Ser Pro Ile His Ser Thr Gly Gln Gln Phe Ile Tyr
                                    460
Ser Gly Cys Ser Thr Gly Lys Val Val Val Tyr Asp Leu Leu Ser
                                    475
Gly His Ile Val Lys Lys Leu Thr Asn His Lys Ala Cys Val Arg
                                    490
Asp Val Ser Trp His Pro Phe Glu Glu Lys Ile Val Ser Ser Ser
                500
Trp Asp Gly Asn Leu Arg Leu Trp Gln Tyr Arg Gln Ala Glu Tyr
                515
                                    520
Phe Gln Asp Asp Met Pro Glu Ser Glu Glu Cys Ala Ser Ala Pro
                                    535
                530
Ala Pro Val Pro Gln Ser Ser Thr Pro Phe Ser Ser Pro Gln
                                    550
                545
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<210> 25
<211> 434
<212> PRT
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<213> Homo sapiens

<220>
<221> misc feature

<223> Incyte ID No: 2226116CD1

<400> 25

 Met
 Arg
 Pro
 Ser
 Ser
 Ser
 Val
 Ser
 Val
 Ser
 Cys
 Pro
 Ala
 Leu
 Asn
 10
 ...
 ...
 ...
 15

 Gln
 Val
 Ser
 His
 Phe
 Ala
 Asn
 Leu
 Thr
 Ser
 Val
 Gly
 Ala
 Met
 Ala

 Pro
 Ala
 Arg
 Cys
 Phe
 Ser
 Ala
 Arg
 Leu
 Arg
 Thr
 Val
 Phe
 Gly
 Gly

 Val
 Gly
 His
 Trp
 Ala
 Leu
 Ser
 Thr
 Trp
 Ala
 Gly
 Leu
 Leu

```
55
                 50
Arg Leu Leu Pro Gln Arg Ala Ser Pro Arg Leu Leu Ser Val Gly
                                    70
                 65
Arg Ala Asp Leu Ala Lys His Gln Glu Leu Pro Gly Lys Lys Leu
Leu Ser Glu Lys Lys Leu Lys Arg Tyr Phe Val Asp Tyr Arg Arg
                                   100
                 95
Val Leu Val Cys Gly Gly Asn Gly Gly Ala Gly Ala Ser Cys Phe
                110
                                    115
His Ser Glu Pro Arg Lys Glu Phe Gly Gly Pro Asp Gly Gly Asp
                125
                                   130
Gly Gly Asn Gly Gly His Val Ile Leu Arg Val Asp Gln Gln Val
                140
                                    145
Lys Ser Leu Ser Ser Val Leu Ser Arg Tyr Gln Gly Phe Ser Gly
Glu Asp Gly Gly Ser Lys Asn Cys Phe Gly Arg Ser Gly Ala Val
                                    175
Leu Tyr Ile Arg Val Pro Val Gly Thr Leu Val Lys Glu Gly Gly
                                    190
Arg Val Val Ala Asp Leu Ser Cys Val Gly Asp Glu Tyr Ile Ala
                                    205
                200
Ala Leu Gly Gly Ala Gly Gly Lys Gly Asn Arg Phe Phe Leu Ala
                                    220
Asn Asn Asn Arg Ala Pro Val Thr Cys Thr Pro Gly Gln Pro Gly
                                   235
                230
Gln Gln Arg Val Leu His Leu Glu Leu Lys Thr Val Ala His Ala
                                    250
                245
Gly Met Val Gly Phe Pro Asn Ala Gly Lys Ser Ser Leu Leu Arg
                                    265
                260
Ala Ile Ser Asn Ala Arg Pro Ala Val Ala Ser Tyr Pro Phe Thr
                                    280
                275
Thr Leu Lys Pro His Val Gly Ile Val His Tyr Glu Gly His Leu
                                    295
                290
Gln Ile Ala Val Ala Asp Ile Pro Gly Ile Ile Arg Gly Ala His
Gln Asn Arg Gly Leu Gly Ser Ala Phe Leu Arg His Ile Glu Arg
                                    325
Cys Arg Phe Leu Leu Phe Val Val Asp Leu Ser Gln Pro Glu Pro
                                    340
Trp Thr Gln Val Asp Asp Leu Lys Tyr Glu Leu Glu Met Tyr Glu
                                    355
                350
Lys Gly Leu Ser Ala Arg Pro His Ala Ile Val Ala Asn Lys Ile
                                   370
Asp Leu Pro Glu Ala Gln Ala Asn Leu Ser Gln Leu Arg Asp His
                380
                                   385
Leu Gly Gln Glu Val Ile Val Leu Ser Ala Leu Thr Gly Glu Asn
                                    400
                395
Leu Glu Gln Leu Leu His Leu Lys Val Leu Tyr Asp Ala Tyr
                                    415
                410
Ala Glu Ala Glu Leu Gly Gln Gly Arg Gln Pro Leu Arg Trp
                                    430
                425
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<210> 26 <211> 826

<212> PRT

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<213> Homo sapiens
<220>
<221> misc feature
<223> Incyte ID No: 2504472CD1
<400> 26
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Asn Arg Val Arg Gly Val Leu Asn Trp Ser Ser Gly Pro Arg Gly
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Leu Leu Ala Phe Gly Thr Ser Cys Ser Val Val Leu Tyr Asp Pro
                 35
Leu Lys Arg Val Val Val Thr Asn Leu Asn Gly His Thr Ala Arg
                 50
Val Asn Cys Ile Gln Trp Ile Cys Lys Gln Asp Gly Ser Pro Ser
                 65
Thr Glu Leu Val Ser Gly Gly Ser Asp Asn Gln Val Ile His Trp
                                     85
Glu Ile Glu Asp Asn Gln Leu Leu Lys Ala Val His Leu Gln Gly
His Glu Gly Pro Val Tyr Ala Val His Ala Val Tyr Gln Arg Arg
                                    115
Thr Ser Asp Pro Ala Leu Cys Thr Leu Ile Val Ser Ala Ala Ala
Asp Ser Ala Val Arg Leu Trp Ser Lys Lys Gly Pro Glu Val Met
                                    145
                140
Cys Leu Gln Thr Leu Asn Phe Gly Asn Gly Phe Ala Leu Ala Leu
                                   160
Cys Leu Ser Phe Leu Pro Asn Thr Asp Val Pro Ile Leu Ala Cys
                                    175
                170
Gly Asn Asp Asp Cys Arg Ile His Ile Phe Ala Gln Gln Asn Asp
                                    190
                185
Gln Phe Gln Lys Val Leu Ser Leu Cys Gly His Glu Asp Trp Ile
                                    205
                200
Arg Gly Val Glu Trp Ala Ala Phe Gly Arg Asp Leu Phe Leu Ala
                                    220
                215
Ser Cys Ser Gln Asp Cys Leu Ile Arg Ile Trp Lys Leu Tyr Ile
                                    235
Lys Ser Thr Ser Leu Glu Thr Gln Asp Asp Asp Asn Ile Arg Leu
Lys Glu Asn Thr Phe Thr Ile Glu Asn Glu Ser Val Lys Ile Ala
                                    265
Phe Ala Val Thr Leu Glu Thr Val Leu Ala Gly His Glu Asn Trp
                275
Val Asn Ala Val His Trp Gln Pro Val Phe Tyr Lys Asp Gly Val
                                    295
Leu Gln Gln Pro Val Arg Leu Leu Ser Ala Ser Met Asp Lys Thr
                                    310
                305
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Ile His Gly Tyr Asp Leu Lys Cys Leu Ala Met Ile Asn Arg Phe
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Gln Phe Val Ser Gly Ala Asp Glu Lys Val Leu Arg Val Phe Ser
Ala Pro Arg Asn Phe Val Glu Asn Phe Cys Ala Ile Thr Gly Gln
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Gly Ala Thr Val Pro Ala Leu Gly Leu Ser Asn Lys Ala Val Phe
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Asn Leu Lys Asn Ile Ser Glu Leu Phe Tyr Tyr Ala Gln Lys Ala
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Val Leu His Pro Thr Gly Pro Leu Tyr Cys Pro Glu Glu Lys Glu
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Met Lys Pro Ala Cys Ile Lys Ala Leu Thr Arg Ile Phe Lys Ile
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Phe Phe Gln Arg Ile Cys Phe Asn Thr Pro Leu Ala Pro Gln Ala
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Leu Glu Asp Val Lys Asn Val Val Arg Lys His Ile Ser Asp Gly
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Val Ala Asp Ser Gly Leu Thr Leu Lys Gly Phe Leu Phe Leu His
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 Lys
 Ser
 Ala
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 Glu
 Pro
 Leu
 Pro
 His
 Gln
 Thr
 Val

 Met
 Arg
 Ile
 Ser
 Ile
 Ser
 Ile
 Ile

85 ...

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Lys	Met	Phe	Gln		Trp	Ser	GIY	Pro		гуѕ	Leu	Den	AId	135
•	~ 1	•	T 7.	125	0	0	~1··	Dwa	130	Gln	Val	Tave	Hie	
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Mob	Gln	17-7	т1.	140	Dro	G1 n	Dhe	Gln		Δsn	Phe	Tle	Ser	
Mec	GIII	val	116	155	FIO		FIIC	0111	160	1102				165
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His	Arg	Ile	Asp		Asn	Trp	Arg	Arg		GIU	Leu	ьуѕ	Ser	255
*	Val	•	*	245	*** -	7	7 am	ui c	250	Tla	Thr	Cve	T.611	
гÀ2	vaı	Leu	гÀг	260	HIS	ASP	Asp	птэ	265	116	1111	Cys	Tie (T	270
Dhe	Cys	Glv	Δen		Tle	Va 1	Ser	Glv		asp	asp	Asn	Thr	
- 110	Cys	- -y		275		• • • • • • • • • • • • • • • • • • • •		1	280	2	•			285
Lvs	Val	Trp	Ser		Val	Thr	Gly	Lys	Cys	Leu	Arg	Thr	Leu	Val
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n ra	Asp	71-	Thr	350	7 ~~	17 a 1	Trn	Δen		Glu	Thr	Glv	Gln	
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Thr	Ser	Glv	Met		Len	Lvs	Asp	Asn		Leu	Val	Ser	Glv	
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نودي . علاجه

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(54) Title: GTPASE ASSOCIATED PROTEINS

(57) Abstract

The invention provides human GTPase associated proteins (GTPAP) and polynucleotides which identify and encode GTPAP. The invention also provides expression vectors, host cells, antibodies, agonist, and antagonists. The invention also provides methods for diagnosing, treating, or preventing disorders associated with expression of GTPAP.

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. CLASSIFICATION OF SUBJECT MATTER PC 7 C12N15/12 C07 A. CLASS CO7K14/47 C07K16/18 A61K38/17 G01N33/68 According to International Patent Classification (IPC) or to both national classification and IPC B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) IPC 7 C07K C12N Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Citation of document, with indication, where appropriate, of the relevant passages Category ° Relevant to claim No. Х MOOSLEHNER K ET AL: "STRUCTURE AND 1-12 EXPRESSION OF A GENE ENCODING A PUTATIVE GTP-BINDING PROTEIN IDENTIFIED BY PROVIRUS INTEGRATION IN A TRANSGENIC MOUSE STRAIN" MOLECULAR AND CELLULAR BIOLOGY 1991, vol. 11, no. 2, 1991, pages 886-893. XP000891270 ISSN: 0270-7306 abstract; figure 1 WO 98 37196 A (LUDWIG INST CANCER RES) A 1-20 27 August 1998 (1998-08-27) abstract; claims 1-52; examples 1-8 WO 94 16069 A (SCHERING CORP ; NAKAFUKU Α 1-6,9-15 MASATO (JP); KAZIRO YOSHITO (JP)) 21 July 1994 (1994-07-21) abstract; claims 1-39 -/--Further documents are listed in the continuation of box C. Patent family members are listed in annex. Special categories of cited documents: T later document published after the international filing date "A" document defining the general state of the art which is not considered to be of particular relevance or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "E" earlier document but published on or after the international *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to filing date "L" document which may throw doubts on priority claim(a) or which is cited to establish the publication date of another citation or other special reason (as specified) involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the "O" document referring to an oral disclosure, use, exhibition or document is combined with one or more other, such doc ments, such combination being obvious to a person skilled in the art. document published prior to the international filing date but later than the priority date claimed "&" document member of the same patent family Date of the actual completion of the international search Date of mailing of the international search report **0** 5. 07. 00 24 March 2000 Name and mailing address of the ISA Authorized officer European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo ni, Gurdjian, D Fax: (+31-70) 340-3016

Intern sal Application No PCT/US 99/28013

ategory °	tion) DOCUMENTS CONSIDERED TO BE RELEVANT	<u> </u>		
alegury	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.		
	WO 91 15582 A (CETUS CORP) 17 October 1991 (1991-10-17) abstract; claims 1-46; example 10	1-16,19, 20		
•	WO 90 00607 A (CETUS CORP) 25 January 1990 (1990-01-25) abstract; claims 1-55; figures 3,4		1-14	
	-			

tional application No. PCT/US 99/28013

Int€

Box i	Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)
This Inte	ernational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. χ	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
•	Although claims 19,20 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. X	Claims Nos.: 17 18 20 because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
	see FURTHER INFORMATION sheet PCT/ISA/210
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This Inte	ernational Searching Authority found multiple inventions in this international application, as follows:
•	
1.	As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this international Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. [X]	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
	claims 1-20 partially
Remark	on Protest The additional search fees were accompanied by the applicant's protest.
	No protest accompanied the payment of additional search fees.

Continuation of Box 1.2

Claims Nos.: 17 18 20

Claims 17,18,20 refer to an antagonist and agonist and the use of antagonist of polypeptide of claim 1 without giving a true technical characterization. Moreover, no such compound is defined in the application. In consequence, he scope of said claims is ambigous and vague, and their subject-matter is not sufficiently disclosed and supported (art.5 and 6 PCT). No search can be carried out for such speculative claims the wording of which, is in fact, a mere recitation of the results to be achieved.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

1. Claims: 1-20 (partially)

A protein with amino acid with seq.id. 1 and corresponding nucleotide sequence with seq.id. 30 , method for detecting a polynucleotide, expression vector ,host cell , method for producing a polypeptide , pharmaceutical composition , antibody , agonist and antagonist , method for preventing a disorder

2. Claims: 1-20 (partially)

A protein with amino acid with seq.id.2 and corresponding nucleotide sequence with seq.id. 31, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

3. Claims: 1-20 (partially)

A protein with amino acid with seq.id.3 and corresponding nucleotide sequence with seq.id. 32, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

4. Claims: 1-20 (partially)

A protein with amino acid with seq.id.4 and corresponding nucleotide sequence with seq.id. 33, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

5. Claims: 1-20 (partially)

A protein with amino acid with seq.id.5 and corresponding nucleotide sequence with seq.id. 34, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

Claims: 1-20 (partially)

A protein with amino acid with seq.id.6 and corresponding nucleotide sequence with seq.id. 35, method for detecting a polynucleotide, expression vector, host cell, method for

producing a polypeptide , pharmaceutical composition , antibody , agonist and antagonist , method for preventing a disorder

7. Claims: 1-20 (partially)

A protein with amino acid with seq.id.7 and corresponding nucleotide sequence with seq.id. 36, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

8. Claims: 1-20 (partially)

A protein with amino acid with seq.id.8 and corresponding nucleotide sequence with seq.id. 37, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

9. Claims: 1-20 (partially)

A protein with amino acid with seq.id.9 and corresponding nucleotide sequence with seq.id. 38, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

10. Claims: 1-20 (partially)

A protein with amino acid with seq.id.10 and corresponding nucleotide sequence with seq.id. 39, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

11. Claims: 1-20 (partially)

A protein with amino acid with seq.id.11 and corresponding nucleotide sequence with seq.id. 40, method for detecting a polynucleotide, expression vector ,host cell , method for producing a polypeptide , pharmaceutical composition , antibody , agonist and antagonist , method for preventing a disorder

12. Claims: 1-20 (partially)

A protein with amino acid with seq.id.12 and corresponding nucleotide sequence with seq.id. 41, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

13. Claims: 1-20 (partially)

A protein with amino acid with seq.id.13 and corresponding nucleotide sequence with seq.id. 42, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

14. Claims: 1-20 (partially)

A protein with amino acid with seq.id.14 and corresponding nucleotide sequence with seq.id. 43, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

15. Claims: 1-20 (partially)

A protein with amino acid with seq.id.15 and corresponding nucleotide sequence with seq.id. 44, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

16. Claims: 1-20 (partially)

A protein with amino acid with seq.id.16 and corresponding nucleotide sequence with seq.id. 45, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

17. Claims: 1-20 (partially)

A protein with amino acid with seq.id.17 and corresponding nucleotide sequence with seq.id. 46, method for detecting a polynucleotide, expression vector, host cell, method for

producing a polypeptide , pharmaceutical composition , antibody , agonist and antagonist , method for preventing a disorder

18. Claims: 1-20 (partially)

A protein with amino acid with seq.id.18 and corresponding nucleotide sequence with seq.id. 47, method for detecting a polynucleotide, expression vector ,host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

19. Claims: 1-20 (partially)

A protein with amino acid with seq.id.19 and corresponding nucleotide sequence with seq.id. 48, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

20. Claims: 1-20 (partially)

A protein with amino acid with seq.id.20 and corresponding nucleotide sequence with seq.id. 49, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharamaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

21. Claims: 1-20 (partially)

A protein with amino acid with seq.id.21 and corresponding nucleotide sequence with seq.id. 50 , method for detecting a polynucleotide, expression vector ,host cell , method for producing a polypeptide , pharmaceutical composition , antibody , agonist and antagonist , method for preventing a disorder

22. Claims: 1-20 (partially)

A protein with amino acid with seq.id.22 and corresponding nucleotide sequence with seq.id. 51, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

23. Claims: 1-20 (partially)

A protein with amino acid with seq.id.23 and corresponding nucleotide sequence with seq.id. 52, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

24. Claims: 1-20 (partially)

A protein with amino acid with seq.id.24 and corresponding nucleotide sequence with seq.id. 53, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

25. Claims: 1-20 (partially)

A protein with amino acid with seq.id.25 and corresponding nucleotide sequence with seq.id. 54, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

26. Claims: 1-20 (partially)

A protein with amino acid with seq.id.26 and corresponding nucleotide sequence with seq.id. 55, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

27. Claims: 1-20 (partially)

A protein with amino acid with seq.id.27 and corresponding nucleotide sequence with seq.id. 56, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

28. Claims: 1-20 (partially)

A protein with amino acid with seq.id.28 and corresponding nucleotide sequence with seq.id. 57, method for detecting a polynucleotide, expression vector, host cell, method for

producing a polypeptide , pharmaceutical composition , antibody , agonist and antagonist , method for preventing a disorder

29. Claims: 1-20 (partially)

A protein with amino acid with seq.id.29 and corresponding nucleotide sequence with seq.id. 58, method for detecting a polynucleotide, expression vector, host cell, method for producing a polypeptide, pharmaceutical composition, antibody, agonist and antagonist, method for preventing a disorder

Ir...rmation on patent family members

Intern: 1al Application No PCT/US 99/28013

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